

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

GJE-30

U.S. APPLICATION NO. (if known, see 37 CFR 1.5)

INTERNATIONAL APPLICATION NO.
PCT/GB97/03015INTERNATIONAL FILING DATE
November 3, 1997PRIORITY DATE CLAIMED
November 1, 1996TITLE OF INVENTION Therapeutic Use of an Agent That Stimulates NO or Prostacyclin
Production and Delivery Device

APPLICANT(S) FOR DO/EO/US

John Francis Martin, Seppo Yla-Herttuala, Stephen George Edward Barker

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application as filed (35 U.S.C. 371(c)(2))
 - a. ☐ is transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US).
6. ☐ A translation of the International Application into English (35 U.S.C. 371(c)(2)).
7. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☒ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☐ have not been made and will not be made.
8. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).
9. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371(c)(4)). (unexecuted)
10. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).

Items 11. to 16. below concern document(s) or information included:

11. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
12. ☐ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
13. ☒ A **FIRST** preliminary amendment.
☐ A **SECOND** or **SUBSEQUENT** preliminary amendment.
14. ☐ A substitute specification.
15. ☐ A change of power of attorney and/or address letter.
16. ☒ Other items or information:
Small Entity Form

17. <input checked="" type="checkbox"/> The following fees are submitted: BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) : Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO and International Search Report not prepared by the EPO or JPO \$970.00 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but International Search Report prepared by the EPO or JPO..... \$840.00 International preliminary examination fee (37 CFR 1.482) not paid to USPTO but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$760.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) but all claims did not satisfy provisions of PCT Article 33(1)-(4) \$670.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(1)-(4) \$96.00 ENTER APPROPRIATE BASIC FEE AMOUNT =				CALCULATIONS PTO USE ONLY																																																									
Surcharge of \$130.00 for furnishing the oath or declaration later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).				\$ 840.00																																																									
<table border="1"> <thead> <tr> <th>CLAIMS</th> <th>NUMBER FILED</th> <th>NUMBER EXTRA</th> <th>RATE</th> </tr> </thead> <tbody> <tr> <td>Total claims</td> <td>38 - 20 =</td> <td>18</td> <td>X \$18.00</td> </tr> <tr> <td>Independent claims</td> <td>5 - 3 =</td> <td>2</td> <td>X \$78.00</td> </tr> <tr> <td colspan="3">MULTIPLE DEPENDENT CLAIM(S) (if applicable)</td> <td>+ \$260.00</td> </tr> <tr> <td colspan="3">TOTAL OF ABOVE CALCULATIONS =</td> <td>\$1,320.00</td> </tr> <tr> <td colspan="3">Reduction of 1/2 for filing by small entity, if applicable. A Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28).</td> <td>\$-660.00</td> </tr> <tr> <td colspan="3">SUBTOTAL =</td> <td>\$ 660.00</td> </tr> <tr> <td colspan="3">Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).</td> <td>\$</td> </tr> <tr> <td colspan="3">TOTAL NATIONAL FEE =</td> <td>\$</td> </tr> <tr> <td colspan="3">Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property</td> <td>\$</td> </tr> <tr> <td colspan="3">TOTAL FEES ENCLOSED =</td> <td>\$ 660.00</td> </tr> <tr> <td colspan="3"></td> <td>Amount to be:</td> </tr> <tr> <td colspan="3"></td> <td>refunded</td> </tr> <tr> <td colspan="3"></td> <td>charged</td> </tr> </tbody> </table>				CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	Total claims	38 - 20 =	18	X \$18.00	Independent claims	5 - 3 =	2	X \$78.00	MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$260.00	TOTAL OF ABOVE CALCULATIONS =			\$1,320.00	Reduction of 1/2 for filing by small entity, if applicable. A Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28).			\$-660.00	SUBTOTAL =			\$ 660.00	Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).			\$	TOTAL NATIONAL FEE =			\$	Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property			\$	TOTAL FEES ENCLOSED =			\$ 660.00				Amount to be:				refunded				charged	\$	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE																																																										
Total claims	38 - 20 =	18	X \$18.00																																																										
Independent claims	5 - 3 =	2	X \$78.00																																																										
MULTIPLE DEPENDENT CLAIM(S) (if applicable)			+ \$260.00																																																										
TOTAL OF ABOVE CALCULATIONS =			\$1,320.00																																																										
Reduction of 1/2 for filing by small entity, if applicable. A Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28).			\$-660.00																																																										
SUBTOTAL =			\$ 660.00																																																										
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).			\$																																																										
TOTAL NATIONAL FEE =			\$																																																										
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property			\$																																																										
TOTAL FEES ENCLOSED =			\$ 660.00																																																										
			Amount to be:																																																										
			refunded																																																										
			charged																																																										

- a. ☐ A check in the amount of \$_____ to cover the above fees is enclosed.
- b. ☒ Please charge my Deposit Account No. 19-0065 in the amount of \$ 660.00 to cover the above fees.
A duplicate copy of this sheet is enclosed.
- c. ☒ The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment to Deposit Account No. 19-0065. A duplicate copy of this sheet is enclosed.

NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.

SEND ALL CORRESPONDENCE TO

Doran R. Pace
Saliwanchik, Lloyd & Saliwanchik
A Professional Association
2421 N.W. 41st Street, Suite A-1
Gainesville, FL 32606-6669

SIGNATURE

Doran R. Pace

NAME _____

38,261

REGISTRATION NUMBER



USA #3

Applicant or Patentee: J.F. MARTIN, S.YLA-HERTTUALA, S.G.E. BARKER Attorneys Docket No. GJE-30

Application or Patent No: 09/297,486

Filed or Issued: April 30, 1999

Title: THERAPEUTIC USE OF AN AGENT THAT STIMULATES NO OR PROSTACYCLIN PRODUCTION AND DELIVERY DEVICE

VERIFIED STATEMENT (DECLARATION) CLAIMING SMALL ENTITY STATUS

(37 CFR 1.9(f) and 1.27(c)) - SMALL BUSINESS CONCERN

I hereby declare that I am

☐ the owner of the small business concern identified below:☒ an official of the small business concern empowered to act on behalf of the concern identified below:

NAME OF CONCERN EUROGENE LIMITED

ADDRESS OF CONCERN Marquis House, 67/68 Jermyn Street, London SW1Y 6NY, United Kingdom.

I hereby declare that the above-identified small business concern qualifies as a small business concern as defined in 13 CFR 121.12, and reproduced in 37 CFR 1.9 (d), for purposes of paying reduced fees to the United States Patent and Trademark Office, in that the number of employees of the concern, including those of its affiliates, does not exceed 500 persons. For purposes of this statement, (1) the number of employees of the business concern is the average over the previous fiscal year of the concern of the persons employed on a full-time, part-time or temporary basis during each of the pay periods of the fiscal year, and (2) concerns are affiliates of each other when either, directly or indirectly, one concern controls or has the power to control the other, or a third party or parties controls or has the power to control both.

I hereby declare that rights under contract or law have been conveyed to and remain with the small business concern identified above with regard to the invention described in

☐ the specification filed herewith.☒ the application identified above.☐ patent identified above.

If the rights held by the above identified small business concern are not exclusive, each individual, concern or organisation having rights in the invention must file separate verified statements averring to their status as small entities, and no rights to the invention are held by any person, other than the inventor, who would not qualify as an independent inventor under 37 CFR 1.9(c) if that person made the invention, or by any concern which would not qualify as small business concern under 37 CFR 1.9(d), or a non profit organisation under 37 CFR 1.9(e)

Each person, concern or organisation having any rights in the invention is listed below:

☒ no such person, concern or organisation exists.☐ each such person, concern or organisation is listed below.

Separate verified statements are required from each named person, concern or organisation having rights to the invention averring to their status as small entities. (37 CFR 1.27)

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28 (b)).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardise the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

NAME OF PERSON SIGNING N R Parker

TITLE OF PERSON OTHER THAN OWNER Director

ADDRESS OF PERSON SIGNING Eurogene Limited, Marquis House,
67/68 Jermyn Street, London SW1Y 6NY, United Kingdom

SIGNATURE X

X DATE 19 May 1999

April 30, 1999

PRELIMINARY AMENDMENT
Patent Application
Docket No. GJE-30

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant(s) : John Francis Martin, Seppo Yla-Herttuala, Stephen George Edward Barker
Docket No. : GJE-30
For : Therapeutic Use of an Agent That Stimulates NO or Prostacyclin Production
and Delivery Device

Box PCT
Assistant Commissioner for Patents
Washington, D.C. 20231

PRELIMINARY AMENDMENT

Sir:

Please amend the above-identified patent application as follows:

In the Claims:

The following amendments are made with respect to the claims in the international application PCT/GB97/03015 after amendements under Article 19 of the PCT and as attached to the International Preliminary Examination Report (IPER). Therefore, please replace existing pages 65 of the international application with the amended claim sheet (replacement page 65) of the annex attached to the IPER, and make the following amendments to the pending claims.

Claim 1 (amended):

[Use of an agent that is an agonist of a receptor to which VEGF binds, or a nucleic acid encoding the agonist, for the manufacture of a medicament] A method for the treatment or prevention of intimal hyperplasia of a blood vessel, where the endothelium is wholly or largely intact, wherein said method comprises administration to a person or animal of an effective amount

of an agent that is an agonist of a receptor to which VEGF binds, or a nucleic acid encoding said agonist.

Claim 2 (amended):

[Use] The method according to claim 1, wherein [the] said blood vessel is an artery.

Claim 3 (amended):

[Use] The method according to claim 1 [or claim 2], for the treatment or prevention of stenosis induced by a surgical procedure or associated with pulmonary artery hypertension.

Claim 4 (amended):

[Use] The method according to claim 3, wherein [the] said surgical procedure is angioplasty, coronary bypass surgery, surgical anastomosis or endarterectomy.

Claim 5 (amended):

[Use according to any preceding claim] The method according to claim 1, for the treatment or prevention of stenosis of the blood vessel.

Claim 6 (amended):

[Use according to any preceding claim] The method according to claim 1, for the treatment or prevention of restenosis of the blood vessel.

Claim 7 (amended):

[Use according to any preceding claim] The method according to claim 1, wherein [the] said agent is a protein having the function of human VEGF, or a nucleic acid encoding [the] said protein.

Claim 8 (amended):

[Use] The method according to claim 7, wherein [the] said protein has the sequence of SEQ. ID No. 2, SEQ. ID No. 4, SEQ. ID No. 6 or SEQ. ID No. 8, or an active fragment thereof.

Claim 9 (amended):

[Use according to any of claims 6 to 8] The method according to claim 1, wherein [the] said agent is a nucleic acid in association with a viral or non-viral vector.

Claim 10 (amended):

An implant for therapeutic use, adapted to be placed at or near the site of hyperplasia to be treated or prevented, and containing an agent as defined in [any preceding claim] claim 1.

Claim 11 (amended):

[An] The implant according to claim 10, which is a silastic implant or a biodegradable implant.

Claim 12 (amended):

[An] The implant according to claim 10 [or 11], which is in the form of a collar for fitting around a blood vessel at or near the site of the hyperplasia to be treated or prevented.

Claim 13 (amended):

[An] The implant according to [claims 10 to 12, having] claim 10, comprising an outer wall substantially impermeable to the agent comprised in it.

Claim 14 (amended):

[Use of an agent as defined in any of claims 6 to 9, for the manufacture of a medicament for] A method of therapy [of] for a condition that can be treated or prevented by stimulation of nitric oxide (NO) and/or prostacyclin production *in vivo*, wherein said method comprises administration to a person or animal of an effective amount of an agent, wherein said agent is a nitric oxide synthase, an agonist of a receptor to which VEGF binds, or a nucleic acid encoding said synthase or said agonist.

Claim 15 (amended):

[Use] The method according to claim 14, wherein the condition is hypertension[, e.g. essential hypertension, primary pulmonary hypertension or cor pulmonale].

Claim 17, line 1: Delete "A" and insert --The--.

Claim 18, line 1: Delete "A" and insert --The--.

Claim 18, line 1: Delete "wherein the" and insert --wherein said--.

Claim 19, line 1: Delete "A" and insert --The--.

Claim 20, line 1: Delete "A device according to any of claims 17 to 19" and insert --The device according to claim 17--.

Claim 20, line 1: Delete "the" and insert --said--.

Claim 21, line 1: Delete "A device according to any of claims 17 to 20, wherein the" and insert --The device according to claim 17, wherein said--.

Claim 22, line 1: Delete "A device according to any of claims 17 to 20, wherein the" and insert --The device according to claim 17, wherein said--.

Claim 23, line 1: Delete "A device according to any of claims 16 to 22, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 24, line 1: Delete "A device according to any of claims 16 to 22, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 25, line 1: Delete "A device according to any of claims 16 to 24, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 26, line 1: Delete "A" and insert --the--.

Claim 26, line 1: Rewrite "claims" as --claim--.

Claim 27, line 1: Delete "A device according to any of claims 16 to 26, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 28, line 1: Delete "A device according to any of claims 16 to 27, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 29, line 1: Delete "A device according to any of claims 16 to 28, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 30, line 1: Delete "A device according to any of claims 16 to 29, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 31, line 1: Delete "A device according to any of claims 16 to 29, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 32, line 1: Delete "A device according to any of claims 16 to 29, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 33, line 1: Delete "A" and insert --The--.

Claim 33, line 1: Delete "or claim 32".

Claim 34, line 1: Delete "A device according to any of claims 16 to 33, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 35, line 1: Delete "A device according to any of claims 16 to 34, wherein the" and insert --The device according to claim 16, wherein said--.

Claim 36, line 1: Delete "any of claims 1 to 10" and insert --claim 1--.

Claim 36, line 2: Delete "any of claims 16 to 35" and insert --claim 16--.

Please add new claims 37 and 38:

1 37. The method according to claim 15, wherein the hypertension condition is
2 selected from the group consisting of essential hypertension, primary pulmonary
3 hypertension and cor pulmonale.

1 38. The device according to claim 32, wherein one body portion is generally arcuate
2 in cross-section transverse to its longitudinal extent so as to enable it to surround the exposed
3 portion of a first blood vessel when that vessel is part-embedded in tissue, and
4 longitudinally-extending edges of the first body portion are arranged to be sealed, in use, to
5 the adventitial wall of the first blood vessel or to adjacent tissue.

The Commissioner is hereby authorized to charge any fees under 37 CFR 1.16 or 1.17 as
required by this paper to Deposit Account 19-0065.

Respectfully submitted,



Doran R. Pace
Patent Attorney
Registration No. 38,261
Phone No.: 352-375-8100
Address : 2421 N.W. 41st Street
Suite A-1
Gainesville, FL 32606

DRP/sl

THERAPEUTIC USE OF GROWTH FACTOR, AND DELIVERY DEVICE,
ESPECIALLY FOR THE TREATMENT OF INTIMAL HYPERPLASIA

Field of the Invention

The present invention relates to the therapeutic use of a growth factor, and
5 particularly to the treatment and prevention of intimal hyperplasia of blood vessels and
other conditions, especially hypertension. The invention relates also to a device that can
be used for delivering the active agent.

Background of the Invention

Intimal hyperplasia is the increase in the number of cells between the endothelium and
10 internal elastic lamina of a blood vessel, particularly in the intimal layer found there, or in
an artery. Intimal hyperplasia is often caused by smooth muscle cell (SMC) proliferation
in the blood vessel wall.

When intimal hyperplasia occurs, *de novo* thickening of the intimal layer or of the
vessel wall, i.e. stenosis, may result. Thus, the blood vessel may become occluded.

15 Also, when an obstruction in a blood vessel has been cleared, intimal hyperplasia
occurring after surgery may lead to the artery's becoming occluded again. This is known
as restenosis.

Proliferation of arterial smooth muscle cells commonly occurs when a blood vessel,
e.g. an artery, is deformed or disturbed during surgery. For example, intimal hyperplasia
20 can lead to *de novo* stenosis following bypass grafts in which a vein is anastomosed to an
artery, and following surgical anastomosis in general. Two examples of surgical
procedures which can give rise to stenosis are coronary bypass grafts and above-knee
femoro-popliteal arterial bypass grafts.

Similarly, restenosis can occur following balloon angioplasty procedures used to clear
25 obstructions in blood vessels, for example balloon angioplasty procedures.

Intimal hyperplasia, whether it leads to stenosis or restenosis, remains a major
problem after various surgical procedures.

Atherosclerotic cardiovascular disease is the leading cause of death in Europe and
North America and prompts a highly significant morbidity consequent upon occlusion of
30 the arterial lumen, either preventing or reducing blood flow, thrombosis superimposed
upon a plaque, with possible distal embolisation, arterial wall weakening, leading to

09/297486-001499

aneurysmal dilation and eventual rupture. Dependant upon site and disease distribution, several options for treatment exist, with arterial bypass grafting the most common surgical intervention. For coronary artery disease, this has now become the most common surgical procedure of all in the United States with >200,000 operations performed each year since
5 1990 and with >20,000 operations performed each year in the United Kingdom. In the aorta, renal, mesenteric and peripheral vessels, the burden of surgical bypass procedures continues to increase, with operation rates in the United States and Europe of 35 - 70 per 100,000 population. In combination, the number of surgical bypass procedures performed each year approximates one million.

10 In the first 24 months following surgery, a very significant number of arterial bypass grafts fail (occlude). Quoted values range from 20% to 30%. This means that for all cardiac and peripheral arterial bypass procedures performed each year in the United Kingdom (approximately 25,000 - 30,000), between 6,000 and 7,000 may be expected to fail, within two years. Failure rates for 're-do' procedures are even higher. Such is the
15 financial cost of failure that, in the United States, it has been calculated that even a modest decrease in failure rates following coronary procedures, from 33% to 25%, might save up to \$750 million from the healthcare budget.

There are three main causes for graft failure within five years from surgery. The first is recognised to occur early, within 30 days of the operation (<5%), and represents
20 technical error (e.g. poor anastomotic technique). Later failure, after 24 months, is generally as a result of progression of the original atherosclerotic process. However, it is those grafts that occlude between one and 24 months that form the majority of failures (<70%). In these cases, it is SMC intimal hyperplasia that is responsible for progressive narrowing, i.e. stenosis, of the arterial lumen, resulting eventually in complete occlusion.
25 Typically, the SMC intimal hyperplasia is sited around the distal arterial anastomosis and the native vessel wall opposite the anastomosis. It is thus a primary pathology at this site and not restenosis at a site of previous intimal hyperplasia as might occur following angioplasty. SMC intimal hyperplasia can occur at the more proximal arterial anastomosis and along the graft itself.

Restenosis after angioplasty can lead to even higher failure rates, from 20 to 50% in the first 6 months following the angioplasty. Stenosis and restenosis both remain major problems after surgery.

To date, numerous methods of treating or preventing intimal hyperplasia have been tested, but none has been clinically satisfactory.

Vascular endothelial growth factor (VEGF) is a naturally-occurring protein. In humans, at least four forms exist, of 121, 165, 189 and 206 amino acids. The cDNA and amino acid sequences of the four forms of human VEGF are given in Houck *et al*, Molecular Endocrinology (1991) vol 5, No. 12, pages 1806-1814. A partial genomic sequence is also given. The cDNA sequence of human VEGF is also given in Leung *et al*, Science (1989) 246:1306-1309, together with the bovine VEGF cDNA sequence.

These four forms are referred to herein as VEGF-121, VEGF-165, VEGF-189 and VEGF-206. It should be understood that this numbering refers to the number of amino acids in the mature protein in each case. The translated protein also includes a 26 amino acid presequence which, in nature, is cleaved during intracellular processing.

VEGF is known to play a role in angiogenesis, where it stimulates the division of vascular endothelial cells (EC), increases endothelial permeability and acts as an endothelial "survival factor" in retinal vessels. For example, VEGF, in the form of recombinant protein or when expressed from a plasmid, can induce the development of new blood vessels when injected intra-arterially into ischaemic limbs. This property has led to its use in repairing arteries whose endothelia have been damaged during surgery. Thus, Asahara *et al*, Circulation (1995) 91: 2793, delivered VEGF, *via* a cannula, to the interior of rat carotid arteries following angioplasty that had denuded the endothelium of the artery; it was found that VEGF stimulated the reendothelialisation of the artery which, in turn, appeared to contribute to suppression of intimal hyperplasia.

VEGF protein and gene are disclosed in WO-A-9013649, and their use for treating trauma of the vascular epithelium, diabetic ulcers and blood vessel wounds is proposed. VEGF fragments are described in WO-A-9102058, and their use in angiogenesis and re-endothelialisation of inner vascular surfaces, e.g. in the treatment of ulcers.

GB-A-2298577 discloses a non-restrictive, porous, external stent for arteriovenous bypass grafting procedures. This stent has beneficial effects on luminal size and on medial and intimal thickening.

WO-A-9423668 discloses a device for the local delivery of an agent into a blood vessel, including a reservoir formed between two elements thereof. Its use requires implantation, i.e. cutting through the vessel and then securing the device to the vessel walls. The device is partially porous. The reservoir is in direct contact with luminal blood flow. This involves the risk of infection.

US-A-3797485 discloses a device for delivering a drug to the adventitial surface of a blood vessel. It is provided with permanent walls and transcutaneous tubes for the delivery of drug in liquid form. The intention is that the drug should pass to another site.

Summary of the Invention

The present invention seeks to treat and/or prevent all of the conditions described above, in as much as they arise from intimal hyperplasia. Surprisingly, properties of VEGF have been identified, indicating that it can be used against intimal hyperplasia in different ways.

A collar was placed around the outside of the artery of a rabbit. This procedure normally causes intimal hyperplasia in the rabbit artery, leading to thickening of the arterial wall, which is similar to the stenosis that can occur in human arteries following bypass operations. When the collar was used to deliver DNA encoding VEGF to the arterial wall using a plasmid/liposome vector, the VEGF gene was overexpressed in the arterial wall, including the endothelial layer. Intimal hyperplasia was inhibited. It has been found that the adventitial collar is suitable for arterial gene transfer with all tested gene delivery systems.

This demonstrates that VEGF, in addition to stimulating reendothelialisation in cases where the endothelium is damaged, is capable of suppressing intimal hyperplasia in situations where intimal hyperplasia arises when the endothelium is wholly or largely intact. Therefore, it is potentially useful not only in suppressing restenosis after angioplasty but in preventing or treating *de novo* stenosis in other surgical situations. There is thus a contrast between the new findings and previous findings, where VEGF was found to

stimulate regrowth, or healing, of the endothelium. It is likely that the new findings arise from a different mechanism of action of VEGF.

Furthermore, the new findings demonstrate that effective agents can be delivered to the exterior of the blood vessel, to treat intimal hyperplasia. This has several advantages.

- 5 In particular, the therapeutic agent is not washed away from the site of the hyperplasia by blood flow as with intraluminal delivery. A delivery reservoir can be maintained around the blood vessel, and there is no need for any intraluminal manipulations which damage the endothelium of the blood vessel (and can themselves trigger intimal hyperplasia).

- 10 More particularly, the present invention enables agents for countering SMC intimal hyperplasia to be applied directly to the adventitial surface of the arterial wall (i.e. closest to those cells in the outer media). Any agent used can be applied specifically at the sites most likely to develop an intimal hyperplastic lesion, since these sites are readily exposed at the time of operation.

- VEGF mediates its known effects *via* specific high-affinity tyrosine kinase receptors
15 flk-1/KDR and flt-1 which are only expressed on EC and monocytes. Without wishing to be bound by theory, it is considered likely that the effects of VEGF in the inhibition of hyperplasia are also mediated through the same receptors. Accordingly, the invention also extends to the use of other agonists of the receptors to which VEGF binds, or other materials having the same mechanism of action, to treat or prevent intimal hyperplasia.
20 The specific location of VEGF receptors also confers an advantage of VEGF as compared to many other growth factors and cytokines suggested for the treatment of intimal thickening; the effects of VEGF are more specific to EC since, in the absence of monocytes, high affinity VEGF receptors in the arterial wall are only expressed on EC.

- For example, it has been found that the mechanism of VEGF's inhibition of intimal
25 hyperplasia in situations where the endothelium is wholly or largely undamaged is at least partly *via* the nitric oxide (NO) pathway, as administration of the NO synthesis inhibitor L-NAME counteracts VEGF's effects on intimal hyperplasia in the collar model described above. Thus, VEGF stimulates NO production.

- It is also possible that VEGF has other biological effects that contribute to its
30 inhibition of intimal hyperplasia. In particular, it has been found that VEGF overexpression stimulates production of prostacyclin, activation of cytosolic phospholipase A₂ and von

Willebrand's factor secretion by EC. It may be the case that VEGF's stimulation of NO production and its stimulation of prostacyclin production act in tandem to suppress intimal hyperplasia.

The finding that VEGF acts to stimulate NO and prostacyclin production also suggests that VEGF and agonists of the receptors to which VEGF binds will be useful in the treatment of other NO-linked and/or prostacyclin-linked conditions. In particular, Forte *et al*, Lancet (1997) 349:837-42, have shown that NO levels are low in individual suffering from hypertension. VEGF may therefore be useful in the treatment of prevention of various forms of hypertension. Similarly, VEGF may be useful in the treatment of atherosclerosis.

According to one aspect of the present invention, an agent that has any of the given characteristics found for VEGF is used for the manufacture of a medicament for the treatment or prevention of intimal hyperplasia, e.g. of stenosis. The agent may be provided in the form of an implant. More particularly, the agent stimulates production of NO or prostacyclin; it may be an agonist of a receptor to which VEGF binds, e.g. VEGF itself or a fragment thereof, or a nucleic acid encoding such an agonist.

According to a second aspect of the invention, a device for use in the delivery of a therapeutic agent to a blood vessel in a patient, comprises a body adapted to provide a seal around the vessel, the agent being held within or associated with the device so that, in use, the agent comes into contact with the adventitial surface of the vessel. Such devices can be biodegradable, and do not require permanent transcutaneous delivery tubes.

Description of the Invention

As suggested above and demonstrated in the Examples, various agents, including nitric oxide synthase and a nucleic acid encoding it, are suitable for use in the invention. The agent will often be described herein as the VEGF protein or nucleic acid, and those references, and references to VEGF itself, are given by way of example. Any of the forms of VEGF described above may be used for the purposes of this invention.

Herein, references to these VEGF protein sequences are to be understood to refer both to sequences comprising the presequence and sequences lacking the presequence. VEGF proteins with and without the presequence are suitable for the practice of the invention. Similarly, references to VEGF nucleic acid (DNA and RNA) sequences relate

to both sequences encoding the presequence and sequences that do not encode the presequence.

It should be noted that Houck *et al*, *supra*, give the sequence of VEGF-165 as including the amino acid asparagine (N or Asn) at position 141 (115 in the notation of Houck *et al*, which corresponds to the mature protein). Houck *et al* give this amino acid as lysine (K or Lys) in VEGF-121, VEGF-189 and VEGF-206, and the cDNA sequence (of VEGF-206) quoted in Houck *et al* supports this. Therefore, in the invention, the amino acid at position 141 may be asparagine (N or Asn) or lysine (Lys or K). Each amino acid is encoded by the appropriate triplet codon in nucleic acid sequences of the invention (for DNA, these codons may be AAA or AAG for lysine and AAT or AAC for asparagine). This applies especially to VEGF-165.

The four forms are encoded by the same gene but generated by alternative splicing at the RNA level. Thus, there is a full-length form of human VEGF and three known truncated forms. VEGF-121 and VEGF-165 are soluble and are secreted forms. Similarly, the 26 amino acid presequence is hydrophobic and is believed to decrease the solubility of the protein. Thus, forms of VEGF without the presequence are preferred, as they are expected to have higher solubility. All forms of VEGF are suitable for the practice of the invention, though secreted forms are preferred. VEGF proteins suitable for the practice of the invention may also originate from other species, although human VEGF is preferred. For example, mouse, rabbit and cow VEGF have been cloned and their sequences are available.

For reference, it should be noted that VEGF-121, 165, 189 and 206 are also referred to in the art as VEGF-120, 164, 188 and 205.

VEGF proteins and nucleic acids (DNA and RNA) are suitable agents for the practice of the invention.

When VEGF protein is used, VEGF protein having the amino acid sequence of SEQ ID No. 2 (VEGF-121), 4 (VEGF-165), 6 (VEGF-189) or 8 (VEGF-206) is preferred. Secreted forms of VEGF are preferred, and so VEGF-121 and VEGF-165 are particularly preferred.

In the practice of the invention, it is preferred to use VEGF DNA encoding VEGF-121, VEGF-165, VEGF-189 or VEGF-206, e.g. having the sequence of SEQ ID No. 1,

3, 5 or 7. DNA sequences encoding secreted forms of human VEGF are preferred. Thus, DNA sequences of SEQ ID No. 1 and 3 are particularly preferred.

However, the VEGF DNA and proteins suitable for the practice of the invention are not limited to those specific sequences. Rather, the invention also provides for the use of
5 other closely related DNA and protein sequences.

DNA sequences of the invention may be related to that of SEQ ID No. 1, 3, 5 or 7 in a number of ways. For example, DNA sequences suitable for the practice of the invention may be degenerate sequences that encode the same protein, the protein of SEQ ID No. 2, 4, 6 or 8.

10 Alternatively, DNA sequences of the invention may be substantially homologous to that of SEQ ID No. 1, 3, 5 or 7, and encode a protein that differs in amino acid sequence from that of SEQ ID No. 2, 4, 6 or 8 but encodes a protein having VEGF activity. Typically, DNA sequences for use in the invention have at least 70%, at least 80%, at least 90%, at least 95%, or at least 99% sequence homology to the sequence of SEQ ID No.
15 1, 3, 5 or 7.

Similarly, VEGF DNA sequences for use in the invention may encode fragments of VEGF that retain VEGF activity. Fragments of interest are, say, at least 15 amino acids long, e.g. up to 40 or more amino acids. Examples of suitable fragments are 20 amino acids long, e.g. the sequences 1-20, 11-30, 21-40, 31-50, 41-60, 51-70, 61-80, 71-90, 81-
20 100, 91-110, 101-120, 111-130, 121-140, 131-150, 141-160, 151-170, 161-180, 171-190, 181-200, 191-210 and 196-215 of the active VEGF protein shown as SEQ ID No.6.

DNA sequences for use in the invention may, for example, be genomic DNAs or cDNAs, or hybrids between genomic DNA and cDNA, or they may be synthetic or semi-synthetic. They may originate from any species, though DNAs encoding human VEGF are
25 preferred. They may be single-stranded or double-stranded. Genomic DNAs encoding the proteins of SEQ ID. No. 2, 4, 6 and 8 are particularly preferred.

DNA sequences for use in the invention may differ from the sequence shown in SEQ ID No. 1, 3, 5 or 7 by the deletion, insertion or substitution of one or more nucleotides, provided that they encode a protein having VEGF activity. Similarly, they may be
30 truncated with respect to SEQ ID No. 1, 3, 5 or 7 or extended by one or more nucleotides provided that they encode a protein having VEGF activity.

RNA sequences are also suitable for the practice of the invention. In particular, the invention provides for the use of the RNA sequences corresponding to those of SEQ ID No. 1, 3, 5 or 7; these are preferred RNA sequences. The invention also provides for the use of RNA sequences that are related to these sequences in any of the ways described above for DNA sequences. RNA sequences for the invention may be single-stranded or double-stranded. RNAs of the invention may be of any origin. For example, they may originate from any species, although RNAs encoding human VEGF, especially human VEGF having the sequence shown in SEQ ID. No. 2, 4, 6 or 8 are preferred. Synthetic DNAs may also be used, as may semi-synthetic RNAs. Further, DNA transcribed from bacterial plasmids *in vivo* or *in vitro* may be used.

It will be appreciated by those of skill in the art that, in RNA sequences suitable for the practice of the invention, the T residues will be replaced by U.

VEGF proteins for use in the invention are encoded by DNA or RNA sequences of the invention as defined above. Preferred proteins of the invention are the proteins of SEQ ID No. 2, 4, 6 and 8 though the invention also provides for the use of other proteins having closely related sequences that differ from those of SEQ ID No. 2, 4, 6 or 8 but have VEGF activity.

According to the invention, insofar as it relates to the treatment or prevention of intimal hyperplasia, VEGF activity is the ability completely or partially to inhibit or prevent intimal hyperplasia in a blood vessel, particularly an artery. Proteins that differ slightly in sequence from naturally-occurring VEGF, as described above, retain this property, although not necessarily to as great an extent as VEGF. Similarly, such proteins may exhibit stronger VEGF activity than naturally-occurring VEGF. The same applies to agonists of VEGF, for example peptide, peptoids or other small molecules.

Insofar as the invention relates to other properties of VEGF, VEGF activity is the ability of molecules other than VEGF to reproduce those properties. For example, insofar as the invention relates to VEGF's activity against NO-linked conditions to stimulate NO production, VEGF activity includes the ability to stimulate NO production. Insofar as the invention relates to VEGF's activity against prostacyclin-linked conditions, VEGF activity includes the ability to stimulate prostacyclin production. VEGF proteins suitable for the practice of the invention also typically exhibit one or more of the biological properties of

VEGF that are already known in the art, such as the ability to promote the proliferation of arterial EC *in vitro* and/or *in vivo* or the ability to bind to the receptors to which VEGF binds and activate them in the manner of VEGF.

VEGF proteins suitable for the practice of the invention may be substantially
5 homologous to the VEGF of SEQ ID No. 2, 4, 6 or 8, typically at least 70%, at least 80%, at least 90%, at least 95%, or at least 99% homologous.

VEGF proteins suitable for the practice of the invention may differ from the sequence shown in SEQ ID No. 2, 4, 6 or 8 by the deletion, insertion or substitution of one or more amino acids, provided that they have VEGF activity. Similarly, they may be truncated by
10 one or more amino acids with respect to SEQ ID No. 2, 4, 6 or 8 or extended with respect to SEQ ID No. 2, 4, 6 or 8 by one or more amino acids, provided that they have VEGF activity. In respect of substitutions, conservative substitutions are preferred. Typically, conservative substitutions are substitutions in which the substituted amino acid is of a similar nature to the one present in naturally-occurring VEGF, for example in terms of
15 charge and/or size and/or polarity and/or hydrophobicity. Similarly, conservative substitutions typically have little or no effect on the VEGF activity of the protein.

VEGF proteins for use in the invention that differ in sequence from naturally-occurring VEGF may be engineered to differ in activity from naturally-occurring VEGF. For example, they may be engineered to have stronger VEGF activity. Such manipulations
20 will typically be carried out at the nucleic acid level using recombinant techniques known in the art.

As an alternative to using a VEGF protein as described above, it is possible to use a VEGF agonist. This applies to all the medical applications described herein, especially the treatment of atherosclerosis.

25 In general, a VEGF agonist is a molecule which binds to a receptor to which VEGF binds and has substantially the same effects, leading to VEGF activity, as described herein. In particular, an agonist may bind to the flk-1/KDR or flt-1 receptors. Agonists of the invention are thus referred to both as agonists of VEGF and of the receptors to which VEGF binds.

30 A VEGF agonist may have any chemical structure. For example, a VEGF agonist may be peptide or polypeptide of, for example, up to 10, up to 20, up to 50 or up to 100

amino acids. An agonist may similarly be a modified peptide, or a peptoid. Any suitable modification may be made, including glycosylation, sulphation, COOH-amidation and acetylation, e.g. N-terminal acetylation. Additionally, or alternatively, modified amino acids and/or L-amino acids may be present.

5 Some preferred agonists are fragments, optionally modified as described above, of VEGF, that have VEGF activity. One particularly preferred agonist fragment of VEGF consists of amino acids 1 to 20 (M...H) of SEQ ID No. 4; this peptide is reported to be an agonist of the Flt-1 receptor in human trophoblast cells, by Ahmed *et al*, Lab. Invest. (1997) 76:779.

10 Additional related agonists may also be derived from the N-terminal region of VEGF. For example, with reference to SEQ ID No. 4, peptide agonists of VEGF may comprise the N-terminus of VEGF (amino acid No. 1) and have the amino acid sequence of VEGF up to an amino acid in the range of 25 to 30, 30 to 40, 40 to 50 or 50 to 100. Similarly, preferred agonists may be derived from the N-terminal region of VEGF but comprise a
15 truncated version of the N-terminus. For example, instead of beginning at amino acid No. 1 in SEQ ID No. 4, they may begin at amino acid No. 2, 3, 4, 5, 6, 7, 8, 9 or 10 of SEQ ID No. 4, and have the amino acid sequence of VEGF up to an amino acid in the range of 25 to 30, 30 to 40, 40 to 50 or 50 to 100.

20 Peptide agonists of the invention may also be derived from other parts of the VEGF sequence. For example, a further preferred peptide agonist is a peptide consisting of amino acids 145 to 169 (R...P) of the VEGF-189 sequence of SEQ ID No. 6.

25 Additional related agonists may also be derived from this region of VEGF. For example, with reference to SEQ ID No. 6, peptide agonists of VEGF may have the amino acid sequence of VEGF from an amino acid in the region of 135 to 155 to an amino acid in the region of 160 to 180. For example, peptide agonists derived from this region may have the sequence of VEGF from an amino acid in the region of 135 to 140, 140 to 145 or 145 to 150 to an amino acid in the region of 160 to 165, 165 to 170 or 170 to 175.

Peptide fragments of VEGF as defined above preferably have a total length of 10 to 20, 20 to 25, 25 to 30, 30 to 40 or 40 to 50 amino acids.

30 Other preferred agonists are fragments of the HIV Tat protein. The HIV Tat protein mimics the agonist actions of VEGF and can stimulate angiogenesis in endothelial cells

acting through the Flk-1/KDR receptor; see Albini *et al*, Oncogene (1996) 12:289-297, and Nature Medicine (1996) 2(12):1321-1375. Thus, peptides derived from the HIV-1 Tat sequence such as the amino acids 46-60 of HIV Tat protein have been shown to stimulate growth and migration of endothelial cells; see Albini *et al*, Oncogene (1996) 12:289-297.

- 5 The peptide consisting of amino acids 41 to 65 of HIV-1 Tat protein is a further preferred peptide agonist of the invention.

Agonists of the invention may also have amino acid sequences that differ from that of naturally-occurring VEGF in any of the ways described above for VEGF proteins, as long as their agonist properties are retained.

- 10 Where the agonists of the invention are peptides, they may be generated *in vivo* from nucleic acid sequences encoding them, in order to effect treatment according to the invention. Thus, agonist-encoding nucleic acids may be delivered by gene therapy, as described herein.

- Alternatively, non-peptide VEGF agonists can be used. For example, small molecules
15 that mimic the shape of the parts of VEGF that interact with its receptors may be used.

- In the practice of the invention, VEGF, a nucleic acid encoding VEGF or a VEGF agonist or nucleic acid encoding a VEGF agonist may be delivered to a blood vessel, preferably an artery in any suitable form. Nucleic acids may be delivered in a "naked" form unassociated with a vector, or by means of a gene therapy vector. It is preferred to deliver
20 them by means of any suitable gene therapy vector. In particular, viral or non-viral vectors may be used.

- Suitable viral vectors include adenoviruses, retroviruses, pseudotyped retroviruses, herpesviruses, vaccinia viruses and baculoviruses. Suitable non-viral vectors include oligonucleotides, plasmids, liposomes, cationic liposomes, pH-sensitive liposomes,
25 liposome-protein complexes, immunoliposomes, liposome-protein-polylysine derivatives, water-oil emulsions, polyethylene imines and dendrimers. Preferred vectors include Moloney murine leukaemia virus (MMLV)-derived retroviruses, pseudotyped vesicular stomatitis virus protein-G (VSV-G)-containing retroviruses, adenoviruses, plasmids and plasmid/liposome complexes.

- 30 Where appropriate, two or more types of vector can be used together. For example, a plasmid vector may be used in conjunction with liposomes. Suitable liposomes include,

for example, those comprising dioleoylphosphatidylethanolamine (DOPE), 3 β -[N-(N',N'-dimethylaminoethane)carbamoyl]cholesterol (DC-Chol), or the positively-charged lipid (N-[1-(2,3-dioleyloxy)propyl]-N,N,N-triethylammonium (DOTMA).

Viral vectors of the invention are preferably disabled, e.g. replication-deficient. That is, they lack one or more functional genes required for their replication, which prevents their uncontrolled proliferation *in vivo* and avoids undesirable side-effects of viral infection. Preferably, all of the viral genome is removed except for the minimum genomic elements required to package the viral genome incorporating the VEGF nucleic acid into the viral coat or capsid. For example, it is desirable to delete all the viral genome except the Long Terminal Repeats (LTRs) and a packaging signal. In the case of adenoviruses, deletions are typically made in the E1 region and optionally in one or more of the E2, E3 and/or E4 regions.

Viruses of the invention may be disabled by any suitable technique. For example, genomic deletions may involve complete removal of genes required for replication, or only partial removal. Complete removal is preferred. In general, preferred deletions are of genes required for early transcription of viral genes.

Replication-competent self-limiting or self-destructing viral vectors may also be used.

In general, the VEGF nucleic acid for use in the invention will be comprised within an expression construct that ensures their expression *in vivo* after they have been delivered to the artery, preferably by a vector as defined above. Such constructs typically comprise a promoter capable of directing the expression of the VEGF nucleic acid (and optionally a regulator of the promoter), a translational start codon and, operably linked to the promoter, the VEGF nucleic acid. Preferably, these components are arranged in a 5'-3' orientation.

The construct may also comprise any other suitable components. For example, the construct may comprise a nucleic acid encoding a signal sequence, so positioned in such a position relative to the VEGF nucleic acid that, when it is translated, it is capable of directing the expressed VEGF protein to a given cell type or cell compartment. Any such signal sequence will typically be positioned immediately 3' or immediately 5' to the VEGF nucleic acid, such that the signal sequence and VEGF protein are translated as a single fusion protein, with the signal sequence at the C- or N-terminus.

The construct may also comprise an enhancer which enhances the degree of expression provided by the promoter. Any enhancer which enhances the expression provided by the selected promoter may be used. For example, in the case of the CMV early gene promoter, the CMV early gene enhancer may be used.

- 5 Optionally, the construct may comprise a transcriptional terminator 3' to the VEGF nucleic acid. Any suitable terminator may be used.

Optionally, the construct may comprise a polyadenylation signal operably linked 3' to the VEGF nucleic acid.

- 10 Optionally, the construct may comprise one or more selectable marker genes, e.g. of antibiotic-resistance, to allow selection of transformed cells in culture. For example, cells may be selected for antibiotic resistance in order.

Optionally, the construct may comprise one or more introns, or other non-coding sequences, for example 3' or 5' to the VEGF nucleic acid.

- 15 Any suitable promoter may be used to control the expression of the nucleic acid of the invention. In general, it is preferred to use a viral promoter or a promoter adapted to function in the species of the subject to be treated. Thus, in the case of a human subject, it is preferred to use viral promoters, especially promoters derived from viruses that infect humans, or promoters derived from human genes. Optionally, a promoter may be used in combination with any suitable enhancer.

- 20 Desirably, a "strong" promoter is used, i.e. one that secures high levels of expression of the VEGF protein of the invention. Promoters that achieve overexpression of the VEGF protein are desirable. Preferred promoters include the cytomegalovirus (CMV) promoter, optionally in combination with the CMV enhancer; the human β -actin promoter; the simian virus 40 (SV40) early gene promoter; the Rous sarcoma virus (RSV) promoter; and the
25 retroviral long terminal repeat (LTR) promoter.

- 30 Promoters, and other construct components, are operably linked to the VEGF nucleic acid. Thus, they are positioned in order that they may exert their effect on expression of the VEGF nucleic acid. For example, in the case of a promoter, the promoter is positioned relative to the VEGF nucleic acid such that it is able to direct expression of the VEGF nucleic acid. Desirably, construct components are positioned to allow them to exert their maximum effect on expression.

The nucleic acids or constructs use in the invention, may be incorporated into viral genomes by any suitable means known in the art. Viral genomes may then be packaged into viral coats or capsids by any suitable procedure. In particular, any suitable packaging cell line may be used to generate viral vectors of the invention. These packaging lines
5 complement the replication-deficient viral genomes of the invention, as they include, typically incorporated into their genomes, the genes which have been deleted from the replication-deficient genome. Thus, the use of packaging lines allows viral vectors of the invention to be generated in culture. Suitable packaging lines include derivatives of PA317 cells, Ψ -2 cells, CRE cells, CRIP cells, E-86-GP cells, Fly cells, line 293 cells and 293GP
10 cells.

In the case of non-viral vectors, nucleic acid may be incorporated into the non-viral vectors by any suitable means known in the art.

As desired, vectors, especially viral vectors, may be selected to achieve integration of the nucleic acid or construct into the genome of the cells of the subject to be treated,
15 or to leave the nucleic acid or construct free in the cytoplasm. Integrative vectors are preferred.

VEGF proteins or VEGF nucleic acids for use in the invention, preferably associated with a viral or non-viral vector, as described above, may be administered to arteries in any suitable manner in order to effect treatment of hyperplasia. For example, VEGF or a
20 nucleic acid encoding VEGF may be administered to the exterior wall of the blood vessel, e.g. artery, or to the blood vessel endothelium, e.g. the arterial endothelium, for example *via* the lumen. Local gene transfer is likely to be advantageous over the administration of recombinant VEGF protein since infused compounds are rapidly flushed away by blood flow and short half-life in blood.

25 Once delivered, VEGF nucleic acids of the invention are expressed to produce VEGF proteins, which in turn effect treatment or prevention of intimal hyperplasia. Expression may take place in any cell type or types in the blood vessel, e.g. arterial, wall.

Preferably, expression occurs in such a location that the expressed VEGF is able to reach the endothelium of the blood vessel, e.g. artery. For example, expression may occur
30 in the smooth muscle cells and/or in the endothelium. Most preferably, expression takes place at least in the endothelium of the blood vessel, e.g. artery.

For example, VEGF protein or nucleic acid may be delivered to the outside of the blood vessel, e.g. artery, by direct injection around the site of the hyperplasia to be treated or prevented, or by injection into the lumen of the blood vessel, e.g. artery.

More preferably, the VEGF protein or nucleic acid is delivered by means of an
5 implant placed externally to the blood vessel, e.g. artery, in proximity to the site of the hyperplasia to be treated. Such an implant contains the VEGF protein or nucleic acid or the vector and provides a reservoir of the agent. The VEGF protein or nucleic acid (preferably in association with a vector) may be introduced into the implant before or after the implant is introduced into the subject to be treated. For example, the implant may be
10 fitted in the vicinity of the blood vessel, with the VEGF protein or nucleic acid being introduced into the implant, e.g. by injection, subsequently.

Preferably, the implant is placed in direct contact with the blood vessel, e.g. artery. This is especially preferred when retroviral vectors are used to deliver VEGF nucleic acids, as the physical distortion of the blood vessel may induce smooth muscle cell proliferation,
15 which increases the efficiency of gene transfer by retroviral vectors. This proliferation, like the proliferation induced by the hyperplasia itself, is overcome or at least ameliorated, by the delivery of the VEGF protein or nucleic acid. Similarly, it is preferred for the implant to be in contact with the artery when employing other vectors that exhibit increased efficiency of gene transfer when their target cells are dividing. For example, cell
20 proliferation may also enhance gene transfer efficiency with plasmid/liposome complexes.

Such implants may be in any suitable form. Preferably, the implant is in the form of a collar which surrounds, partially or completely, preferably completely, the artery, at or near the site of the hyperplasia to be treated or prevented.

Extravascular gene delivery avoids procedures such as balloon catheterization or high
25 pressure fluid which may lead to endothelial damage or denudation. Transfected genes are preferably applied *via* a silastic or biodegradable implant, preferably a collar placed next to, preferably around, the outside of the blood vessel. The endothelium suffers little or no damage. This is a major advantage of this form of delivery.

When, according to the invention, vectors are applied directly on the adventitial
30 surface of a blood vessel within a collar, close contact with the adventitia is maintained. In rabbit arteries, a collar alone typically leads to the formation of a neointima within 7-14

days after the operation. The collar also maintains a high concentration of vector at the adventitial surface.

Implants, preferably collars, may be made of any suitable material. Silastic implants, i.e. implants comprising silicone rubbers, are one preferred alternative. Most preferred are
5 biodegradable implants. Any suitable biodegradable material may be used.

Within the implant, e.g. collar, the VEGF protein or nucleic acid may be contained in any way. Preferably, the structure of the implant, e.g. collar, is such that the VEGF protein or nucleic acid is held in direct contact with the blood vessel wall. Thus, in one embodiment, the structure of the implant leaves a space between the blood vessel wall and
10 the wall of the implant. In the case of a collar, the implant thus forms a hollow container around the blood vessel. Into this space, VEGF nucleic acids or proteins can be introduced, such that they are in contact with the blood vessel wall. Preferably, the extremities of the implant are in contact with the blood vessel wall, thus preventing the escape of the VEGF nucleic acid or protein. Preferably, the outer wall of the collar is
15 impermeable, or substantially impermeable, to the VEGF nucleic acid or protein, thus preventing, or at least limiting, its escape into the surrounding tissue and ensuring its delivery to the blood vessel.

Optionally, the space containing the VEGF nucleic acid or protein may be separated from the wall of the blood vessel by one or more layers of material permeable or semi-
20 permeable to the VEGF or nucleic acid. This may be desirable if gradual delivery is intended and is desired to limit the rate at which VEGF protein or nucleic acid is delivered to the blood vessel wall.

Optionally, the implant, e.g. collar, may be designed to act as an osmotic pump.

Optionally, the VEGF may be contained within a medium within the collar, e.g. a
25 solid or gel medium. This may help to prevent the VEGF protein or nucleic acid escaping into the tissue. In this case, the outer wall of the collar may not need to be in contact with the blood vessel of the extremity of the implant.

Alternatively, the VEGF nucleic acid or protein may be coated onto the surface of the implant which is in contact with the blood vessel in use. Alternatively, the VEGF
30 nucleic acid or protein may be dispersed throughout the structure of the implant.

Some advantages of the use of implants in this way, especially collars, are: (i) they provide a delivery reservoir, allowing for sustained delivery; (ii) no intraluminal manipulations are required and the arterial endothelium remains intact; and (iii) the distortion (e.g. constriction in the case of a collar) created by the implant may enhance the efficiency of gene delivery, as explained above.

A device of the invention generally comprises a body including at least a first substantially impermeable body portion which is shaped to in use extend longitudinally along and at least partially surround a first blood-carrying vessel, the first body portion including longitudinally spaced apart seal portions adapted to seal in use against the adventitial surface of the first blood-carrying vessel, and an intermediate portion between the seal portions which is adapted to in use contain and deliver the at least one agent to the adventitial surface of the first blood-carrying vessel.

Preferred embodiments of devices in accordance with the present invention will now be described, by way of example only, with reference to the accompanying drawings, in which:

Figure 1 is a schematic, longitudinal sectional view of a device of the invention in place around a blood vessel;

Figure 2 is a schematic, coronal sectional view of the same embodiment, along the line A-A shown in Figure 1;

Figure 3 shows in perspective a schematic view of a second embodiment of a device positioned around an end-to-end anastomosis;

Figure 4 is a longitudinal cross-sectional view in the vertical plane of the embodiment of Figure 3;

Figure 5 is a similar view to that of Figure 4, but showing a modified form of that embodiment; with an alternative construction for its seal portions;

Figure 6 shows in perspective a schematic view of a third embodiment of a device positioned around an end-to-side anastomosis;

Figure 7 is a longitudinal cross-sectional view in the vertical plane of the embodiment of Figure 6;

Figure 8 shows in perspective a schematic view of a fourth embodiment of a device positioned around a side-to-side anastomosis;

Figure 9 is a longitudinal cross-sectional view in the vertical plane of the embodiment of Figure 8;

Figure 10 shows in perspective a schematic view of a fifth embodiment of a device positioned around a side-to-side anastomosis, showing a part-embedded blood vessel; and

5 Figure 11 is a longitudinal cross-sectional view in the vertical plane of the embodiment of Figure 10.

By way of background, arterial bypass grafts are commonly used to restore or improve the blood flow to tissues when the native vessels are occluded or significantly stenosed, usually by atheroma. Whether using autologous vein or artery, or a synthetic
10 material such as Dacron or PTFE, grafts are commonly anastomosed to the native vessels in one of three ways: "end-to-end" (Figure 4); "end-to-side" (Figure 7); or "side-to-side" (Figure 9). Of these techniques, end-to-side and side-to-side are much more common than end-to-end. The direction of blood flow is represented by the arrows.

Figures 1 and 2 show blood 1 within a vessel wall 2, and an adventitial collar
15 including a void space 3 defined by a wall 4, e.g. of a biodegradable material. A collar 5 touches the vessel wall at the collar's extremities. This embodiment may be used in the same manner as is described below, for other embodiments.

The embodiment illustrated in Figures 3 and 4 is shown applied to an end-to-end anastomosis. The device comprises a generally tubular body 12 which, in use, extends
20 longitudinally along and surrounds the blood-carrying vessel made up by the end-to-end anastomosis of a graft 13 to a native blood vessel 14.

As is most clearly visible from Figure 4, the body 12 of the device has longitudinally spaced-apart seal portions 15 provided at its opposite ends. When, in use, the device is positioned over the site 16 of the end-to-end anastomosis, these seal portions 15 seal
25 against the adventitial surface of the graft 13 and native blood vessel 14. The radial thickness of the material of the body 12 is greater at the seal portions 15 than it is in an intermediate portion 17. Consequently, when the seal portions 15 seal against the adventitial surfaces of the graft 13 and vessel 14, a space is formed between the interior of the body 12 of the device and the adventitial surfaces of the enclosed ends of the graft
30 13 and vessel 14. This space constitutes a sealed reservoir 18 and is shown longitudinally aligned with the anastomosis 16.

This reservoir enables a pharmaceutical formulation containing one or more agents to be placed in contact with the adventitial surface of the graft and vessels 13,14 at the site of the anastomosis 16. Where the formulation is in the form of a fluid or gel it can for example be injected using a hypodermic needle and syringe, through the wall of the body 12, into the sealed reservoir 18. The agents contained in the formulation advantageously have an anti-proliferative effect, to counter smooth muscle cell intimal hyperplasia at the site of the anastomosis 16 and areas contiguous to it.

The pharmaceutical formulation need not be in fluid or gel form, for example it may be a runny paste having a consistency similar to that of toothpaste. It then remains in contact with the pulsing adventitial surface to which it is exposed, constricting the vessel.

To position the device over the site of the anastomosis 16, the cylindrical body 12 may be slid axially over one of the graft 13 and blood vessel 14 prior to their being anastomosed. The surgeon can then join the graft 13 and vessel 14 together at the anastomosis site 16 and the body 12 may then be slid back over the anastomosis site 16 to occupy the position shown in Figure 4, to enable sealing of the seal portions 15 to the respective adventitial surfaces.

Alternatively, the body 12 of the device may, as shown, be provided with a longitudinal slit 19 along its full length. In this way, the surgeon can anastomose the graft 13 and vessel 14 without introducing the device 12 to the patient's body. Once the anastomosis has been successfully completed, the surgeon can then select an appropriately sized body 12 and apply it around the anastomosed graft and vessel by opening the flexible body 12 along slit 19, slipping it over the anastomosed graft and vessel 13,14 and then sealing the opposed longitudinal edges of the body at the slit 19 together, for example using a conventional "tissue glue", such as the thrombin glue sold under the name Tisseal, or a cyanomethacrylate-based glue.

To concentrate the effect of the pharmaceutical formulation contained within the reservoir 18, and to avoid leakage of its agents to the surrounding tissue, the body 12 is substantially impermeable to the formulation. The material is also, advantageously, biodegradable over a set time course, for example a period of 1 to 5 days, by which time the active agents in the formulation are likely to have become exhausted. The material is also chosen so as not to promote too severe a reaction from the surrounding tissue.

Examples of suitable materials for the body include gelatin, alginate or collagen. These materials also allow the body flexibility and enable the device to be manufactured by molding or extrusion.

5 The wall material of the body 12 may also advantageously be self-sealing, so as to preserve the integrity of the sealed reservoir 18 if it is required to be punctured by a hypodermic needle. Alternatively or additionally, any leak in the wall that is revealed after removal of the needle may be sealed with "tissue glue" or the like.

10 A range of differently sized bodies 12 may be provided to be fitted over differently sized vessels. Lower limb vessels commonly have an external diameter of approximately 6-8 mm. Coronary vessels commonly have an external diameter of approximately 3-5 mm. Accordingly, a range of body sizes of between about 3-10 mm diameter may be made available in sterilised packets to the surgeon. In addition, the size of the body may be varied to influence the volume of the reservoir 18. A suitable size for the reservoir 18 is up to 10 ml, preferably 2-5 ml.

15 To accommodate expansion of the blood-carrying vessel 13,14 caused by pulsatile blood flow therealong, at least the seal portions 15 of the body are advantageously capable of being stretched so as to accommodate expansion of the vessel walls. It is highly desirable to avoid constriction of the vessel walls by the device, whilst at the same time maintaining the seals intact.

20 Figure 5 illustrates different seal portions. The radial thickness of the material of the body 12 is constant along the length of the body, and the intermediate portion 17 is ballooned internally relative to the internal diameter of the body 12 at the seal portions 15. Both seal portions 15 are formed with tails which extend in the longitudinal direction, for example each to contact the respective adventitial surfaces of the graft 13 and vessel 14
25 over a length "X" in the axial direction, of approximately 8-15 mm. These long tails for the seal portions 15 can be made to act in the manner of "flap valves" to help seal the reservoir 18, although clearly no flow through the "flap valve" is desired. Alternatively, the tails may be folded inwardly (not shown) to double the thickness of the body at its ends to form seal portions of greater radial body thickness than the body thickness in the
30 intermediate portion, in a fashion similar to that shown in Figure 4.

To form or help form fluid-tight seals, the surgeon may adhere the seal portions 15 to the adventitial surfaces, for example using a glue of the type mentioned above, for example a "tissue glue". This may not, however, be essential. For example, if the size of the body 12 at the seal portions 15 is chosen to match the girth of the vessels 13,14, it may not be necessary to use any glue. Instead, the surgeon might rely on the radial interference between the internal diameter of the body 12 at the seal portions 15 and the diameter of the adventitial surfaces. This is particularly so with the long-tailed seal portion embodiment of Figure 5. The seal portions should not, however, be so tight on the vessel as to constrict it.

Figures 6 and 7 illustrate an embodiment of the device that is used in conjunction with an end-to-side anastomosis. These drawings show a body 20 having a first body portion 21 and a second body portion 22 branched thereto at an angle of less than 90° to form a generally Y-shaped body. It will be appreciated that the first and second body portions 21,22 may be branched to one another at other branch angles up to and including 90°, in which latter case the body be generally T-shaped. The surgeon may advantageously have available to him a series of differently sized and differently shaped bodies from which he may choose the device most appropriate to the layout and size of the anastomosed vessels. The first body portion 21 may, for example, be approximately 1-10 cm in length; the second body portion 22 may be 1-5 cm in length.

The first body portion 21 is generally tubular and is shown as surrounding a native blood vessel 23. The second body portion 22 is also generally tubular and is shown as surrounding a graft 24 anastomosed to the native blood vessel 23 in end-to-side fashion at anastomosis site 29. As can be seen from Figure 7, the opposite ends of the first body portion 21 are provided with seal portions 25 and the extreme end of the second body portion 22 is provided with a seal portion 26. As in the earlier embodiment, these seal portions 25,26 may advantageously be sealed to the adventitial surfaces of the blood vessel 23 and graft 24 respectively using a "tissue glue".

Figure 7 shows a sealed reservoir 27 formed between the interior of the first body portion 21 and the adventitial surface of the blood vessel 23. The reservoir 27 extends into the interior of the second body portion 22. This sealed reservoir 27 is thus aligned with the site 29 of the anastomosis. The reservoir 27 may advantageously be injected with

a liquid pharmaceutical formulation containing active agents, using a hypodermic needle and syringe.

To facilitate fitment of the device to the vessel 23 and graft 24, the body 20 of the device may be provided to the surgeon in a sterilised package containing two symmetrical halves, split in and symmetrical about the plane of the section illustrated in Figure 7. In such a case, the surgeon would be required to assemble the two identical body halves together after having anastomosed the graft 24 to the vessel 23, and to seal the facing edges of the identical halves together, for example using glue as previously described.

Alternatively, only the first body portion 21 may be provided with a longitudinal slit 28, as shown in Figure 6. In this way the surgeon could slide the second body portion 22 over the end of the graft 24. The surgeon would then be able to anastomose the free end of the graft 24 to the blood vessel 23 and then slide the second body portion 22 back down the graft 24 to cover the site of the anastomosis 29, using the slit 28 to feed the blood vessel 23 into the centre of the first body portion 21 to be surrounded thereby. The surgeon could then seal to one another the facing longitudinal edges of the first body portion 21 at the slit 28 to form the sealed reservoir 27 therein.

It will be appreciated that other configurations may be used for the body portions 21 and 22. For example, the first and second body portions 21 and 22 could be provided separately from one another and be secured to one another to form the sealed reservoir 27 only when *in situ* in the patient.

Figures 8 and 9 illustrate an embodiment of device suitable for use in the situation of a side-to-side anastomosis. The body 30 of the device is shown as comprising a first body portion 31, from which are branched second and third body portions 32,33. The branching forms a generally X-shaped body as shown in Figure 8.

All three body portions 31,32,33 are generally tubular in shape. The device is generally similar to that illustrated in Figures 6 and 7, save for the additional third body portion 33.

First body portion 31 surrounds the occluded native blood vessel 34 and is sealed to the adventitial surface thereof by seal portions 35. Second and third body portions 32,33 surround the graft 36 and are sealed to the adventitial surface of the graft at respective seal portions 37,38. As is most clearly shown in Figure 9, the effect of the seal portions

35,37,38 is to form a sealed reservoir 39 between the interior of the body 30 and the adventitial surfaces of the enclosed vessels, which reservoir 39 can be at least part-filled with a pharmaceutical formulation in the manner described earlier.

To facilitate fitment of the device illustrated in Figures 8 and 9, the device is advantageously provided to the surgeon in at least two parts. For example, the first body portion 31 may be provided separately from a second component comprising second and third body portions 32,33. By providing the body portions with longitudinal slits (not shown), the body portions may be fitted to surround the vessel 34 and graft 36 and then be sealed along those longitudinal slits and sealed to one another along a line of contact, to provide the sealed reservoir 39 around the point of anastomosis 40.

Figures 10 and 11 show a variant of the device shown in Figures 8 and 9 (and use the same reference numerals for common parts). One particularly suitable use for the device of the present invention is in coronary artery bypass graft surgery. In such a situation, the first blood vessel 34 may, as shown, be a coronary artery that is part-embedded in the heart wall 50. A device of the form shown in Figures 8 and 9 could not be fitted to a part-embedded coronary artery 34, as the first body portion would be unable to extend fully around the artery 34. Accordingly, in the embodiments of Figs. 10 and 11, the first body portion 51 of the body 31 does not describe a full circle when viewed in cross-section transverse to its longitudinal extent; instead, it is generally arcuate. In the illustrated embodiment it describes an arc of approximately 180°. This enables the first body portion 51 to be fitted over only the exposed portion of the part-embedded coronary artery 34, to surround it only partly. In this arrangement, the longitudinally-extending edges 41 of the first body portion 51 are sealed by the surgeon either to the adventitial wall of the coronary artery 34 or, as shown, to the surface of the heart wall 50, for example by using tissue glue.

The device of Figures 10 and 11 is also applicable to other surgical procedures in which the first blood-carrying vessel is an artery part-embedded in a wall of the organ supplied by that artery. Such organs include the brain, bladder and uterus.

Although the illustrated embodiments have concentrated on the use of the device to deliver agents to blood-carrying vessels at sites of anastomosis as well as to sites contiguous herewith, the invention is not limited to such uses. The device may, for

example, be used more generally to deliver agents to the adventitial surface of non-anastomosed blood-carrying vessels. For example, following a balloon angioplasty, a device of the form shown in Figures 3 to 5 may be placed around the exterior of an artery in the region of the site of the balloon angioplasty, so as to deliver one or more agents thereto *via* the adventitial surface of the artery.

In the embodiments described above, the sealed reservoirs are shown as taking the form of a radial space or clearance between the adventitial surface of the blood-carrying vessel and the interior of at least the first body portion, which space is at least part-filled in use by a pharmaceutical formulation. Such a space is not, however, essential. For example, in an alternative embodiment, the body may have a generally impermeable flexible outer layer and a flexible inner layer which is impregnated with the formulation and which is arranged to be in contact the adventitial surface in use.

The outer layer may, for example, be made of solid collagen and the inner layer made of sponge-like collagen cross-linked thereto, the sponge-like layer being capable of being impregnated with the pharmaceutical formulation containing the agent to be delivered. In such a situation, the device may be provided to the surgeon for fitment with the formulation already impregnated therein, or it may be wetted with the formulation after fitment, for example by being injected as described earlier.

Alternatively, the agent may be coated onto an internal surface of the body, which surface is just in contact with the blood vessel in use. Alternatively, agent may be dispersed throughout the structure of the body.

It is desirable that the body of the device should have sufficient strength to resist torsional forces. For this purpose, the body may be formed with, for example, an inner layer, e.g. a collagen film, or longitudinal, transverse or helical ribs. Ribs may be provided that subdivide the reservoir into compartments, and to provide additional stability.

The proteins or nucleic acids may be used for the treatment or prevention of intimal hyperplasia arising from any clinical circumstances. For example, it is possible to treat hyperplasia arising after any type of surgical procedure, including angioplasty, for example balloon angioplasty; bypass surgery, such as coronary bypass surgery in which a vein is anastomosed to an artery; other anastomosis procedures, for example anastomosis in the legs; and endarterectomy, for example carotid artery endarterectomy. It is also possible

to treat intimal hyperplasia associated with arterial damage or hypertension, for example pulmonary artery hypertension. The invention provides for treatment of intimal hyperplasia in any type of blood vessel, e.g. in an artery or vein, preferably an artery.

According to the invention, it is possible to treat or ameliorate established intimal hyperplasia or to prevent intimal hyperplasia from arising. Similarly, it is possible to diminish the likelihood of intimal hyperplasia arising, or to diminish the severity of established intimal hyperplasia or hyperplasia that is likely to arise. Treatment according to the invention may take place before, during, or after a surgical procedure, for example in order to reduce the chance of hyperplasia arising after the procedure.

Preferably, the VEGF nucleic acid or protein is administered with a view to preventing or treating *de novo* stenosis. It can, however, also be used to treat or prevent restenosis.

The proteins or nucleic acids of the invention are preferably delivered in the form of a pharmaceutical formulation comprising a pharmaceutically acceptable carrier. Any suitable pharmaceutical formulation may be used.

For example, suitable formulations may include aqueous and non-aqueous sterile injection solutions which may contain anti-oxidants, buffers, bacteriostats, bactericidal antibiotics and solutes which render the formulation isotonic with the blood of the intended recipient; and aqueous and non-aqueous sterile suspensions which may include suspending agents and thickening agents. The formulations may be presented in unit-dose or multi-dose containers, for example sealed ampoules and vials, and may be stored in a frozen or freeze-dried (lyophilized) condition requiring only the addition of the sterile liquid carrier, for example water for injection, immediately prior to use.

It should be understood that, in addition to the ingredients particularly mentioned above, formulations of this invention may include other agents conventional in the art having regard to the type of formulation in question. Of the possible formulations, sterile pyrogen-free aqueous and non-aqueous solutions are preferred.

The proteins, nucleic acids and vectors may be delivered in any suitable dosage, and using any suitable dosage regime. Those of skill in the art will appreciate that the dosage amount and regime may be adapted to ensure optimal treatment of the particular condition

to be treated, depending on numerous factors. Some such factors may be the age, sex and clinical condition of the subject to be treated.

For the delivery of naked nucleic acids encoding VEGF or constructs comprising such nucleic acids, typical doses are from 0.1-5000 μg , for example 50-2000 μg , such as
5 50-100 μg , 100-500 μg or 500-2000 μg per dose. For the delivery of VEGF protein, suitable doses include doses of from 1 to 1000 μg for example from 1 to 10 μg , from 10 to 100 μg , from 100 to 500 μg or from 500 to 1000 μg .

The dosage used for the delivery of VEGF nucleic acids by means of viral or non-viral vectors will depend on many factors, including the efficiency with which the vectors
10 deliver VEGF nucleic acids to cells, and the efficiency with which the VEGF nucleic acids are expressed in the cells.

For example, viral vectors may be delivered in doses of from 10^4 to 10^{14} cfu or pfu/ml, for example 10^4 to 10^6 , 10^6 to 10^8 , 10^8 to 10^{10} , 10^{10} to 10^{12} or 10^{12} to 10^{14} cfu or pfu/ml. Doses in the region of 10^5 to 10^9 cfu or pfu/ml are preferred. The term pfu
15 (plaque-forming unit) applies to certain viruses, including adenoviruses, and corresponds to the infectivity of a virus solution, and is determined by infection of an appropriate cell culture, and measurement, generally after 48 hours, of the number of plaques of infected cells. The term cfu (colony-forming unit) applies to other viruses, including retroviruses, and is determined by means known in the art generally following 14 days incubation with
20 a selectable marker. The techniques for determining the cfu or pfu titre of a viral solution are well known in the art.

For retroviruses, dosages in the region of 10^5 to 10^6 cfu/ml are particularly preferred. For pseudotyped retroviruses, dosages in the region of 10^7 cfu/ml are particularly preferred. For adenoviruses, dosages in the region of 10^9 pfu/ml are particularly preferred.

25 Similarly, such doses may be included within implants of the invention for gradual delivery.

VEGF nucleic acids associated with non-viral vectors may also be delivered in any suitable dosage, by any means of administration, as described above, or gradually from an implant. Suitable doses are typically from 0.1 to 1000 μg of nucleic acid, for example 1
30 to 100 μg , 100 to 500 μg or 500 to 1000 μg , 1000 to 2000 μg , 2000 to 3000 μg or 3000 to 5000 μg . Preferred doses are in the region of 5 to 50 μg , for example 10 to 20 μg .

Dosage schedules will also vary according to, for example, the route of administration, the species of the recipient and the condition of the recipient. However, single doses and multiple doses spread over periods of days, weeks or months are envisaged. Also, as explained above, the delivery of VEGF proteins and nucleic acids may be effected by means of an implant suitable for fitting around a blood vessel, preferably an artery; preferably, the implant is in the form of a collar. Such an implant will effect gradual delivery. For example, delivery may take place over a period of hours, days, weeks or months.

Proteins and nucleic acids of the invention may be administered by any form of administration, for example topical, cutaneous, parenteral, intramuscular, subcutaneous or transdermal administration, or by direct injection into the bloodstream, direct injection into or around the arterial wall or by direct application to mucosal tissues. Preferably, administration is by means of an implant, as described above.

The proteins, nucleic acids and vectors of the invention may be used to treat intimal hyperplasia in any mammal. Treatment of human patients is preferred.

The invention also provides kits for the treatment or prevention of intimal hyperplasia. These kits comprise (i) as an agent, VEGF protein or nucleic acid, preferably in association with a vector, as defined above; and (ii) an implant of the invention in the form of a collar into which the VEGF protein or nucleic acid may be introduced. Preferably, the VEGF nucleic acid or protein is provided in the form of a pharmaceutical formulation comprising a pharmaceutically-acceptable carrier as defined above. Components (i) and (ii) may be packaged in any suitable way. Other components known in the art may also be included, for example standard reagents and/or solutions and/or equipment.

The invention also provides methods of treating or preventing intimal hyperplasia comprising administering to a patient in need of such treatment an effective non-toxic amount of a VEGF protein, nucleic acid or agonist of the invention. Such treatment is effected in the manner described herein.

The implants of the invention, especially implants in the form of collars, as defined above, can also be used for the delivery of agents other than VEGF to blood vessels, e.g.

arteries. Any suitable agent may be delivered in this way, to achieve any desired therapeutic goal.

It has been observed that plasmid/liposome complexes, MMLV retroviruses, VSV-G retroviruses and adenoviruses lead to expression in collared arteries. Gene transfer efficiency was highest with adenoviruses and pseudotyped VSV-G retroviruses also produced a relatively high transfection efficiency. The utility of the replication-deficient VSV-G retroviruses in arterial gene transfer has not been previously demonstrated. Expression was seen in some endothelial cells of the adenovirus-transfected arteries. Since penetration from the adventitia to the intima had occurred, these results raise the general possibility of altering endothelial function in human disease by extraluminal gene transfer using genes other than VEGF. This may also be useful for the expression in the adventitia and outer media of diffusible or secreted gene products which then act elsewhere in the arterial wall. Preferably, such delivery effects treatment of intimal hyperplasia, as defined above, although it may also effect additional or alternative therapeutic goals.

Preferred therapeutic agents for delivery in this manner include proteins other than VEGF that stimulate nitric oxide (NO) production in the arterial wall. The delivery of NO synthases, especially inducible NO synthase (iNOS), to effect treatment or prevention of intimal hyperplasia is particularly preferred.

Other preferred therapeutic agents include agonists which activate the endothelial VEGF receptor (see above). These agonists will typically be small synthetic molecules. Peptides including peptide fragments of VEGF proteins may also be used. In this case, treatment may be effected by delivery of the peptides themselves or of nucleic acids encoding them, as described above for VEGF. These are preferably delivered by the same route, i.e. *via* an implant as defined herein, though they could be delivered systemically.

Preferably, the therapeutic agents will be in the form of nucleic acid encoding a pharmaceutically-active polypeptide or protein. More preferably, this nucleic acid will be comprised within a construct, as defined above. Still more preferably, the nucleic acid or construct will be delivered to the artery by means of a vector as defined above, for example a viral or non-viral vector as defined above.

Thus, extra-arterial gene transfer can be used for the delivery of genetic material into the wall of blood vessels, preferably arteries. From the Examples, it can be seen that changes in medial SMC and even endothelium change can be achieved from the adventitial side, allowing the development of new methods for the treatment of blood vessel, e.g. arterial, disease.

Accordingly, the invention provides the use, in the manufacture of a medicament for the treatment or prevention of intimal hyperplasia of a blood vessel, of NOS (optionally iNOS), or a nucleic acid encoding NOS (optionally iNOS); wherein the NOS protein or nucleic acid is provided in an implant, preferably a collar, as defined above for VEGF.

The invention also provides kits comprising (i) NOS (optionally iNOS) protein or nucleic acid; and (ii) an implant of the invention. These kits are as described above for VEGF. These are suitable for the treatment or prevention of intimal hyperplasia of a blood vessel.

The invention also provides a method of treating intimal hyperplasia of a blood vessel comprising implanting an implant of the invention comprising NOS protein or nucleic acid in the vicinity of the hyperplasia to be treated or prevented, thereby to effect delivery of NOS protein or nucleic acid.

NOS nucleic acid is preferably associated with a vector, as described above for VEGF. Treatment is carried out as described above for VEGF, and dosages and pharmaceutical formulations are also as described above for VEGF.

The invention also provides implants of the invention which comprise NOS (optionally iNOS), protein or nucleic acid.

The finding that VEGF stimulates NO and prostacyclin production in the arterial wall also suggests that VEGF and agonists of VEGF receptors will be useful in the treatment of other NO-linked and/or prostacyclin-linked conditions.

In particular, VEGF and agonists of the receptors to which VEGF binds may be used to treat hypertension, or high blood pressure. Forte *et al*, Lancet (1997) 349:837-42, found that low NO levels are characteristic of individuals suffering from essential (i.e. systemic) hypertension. NO is known to relax the walls of blood vessels; Forte *et al* suggest that impaired NO production reduces this relaxation, leading to constriction of

blood vessels and so to increased blood pressure. Further, in individuals suffering from essential hypertension, levels of prostacyclin may be depressed.

Since VEGF stimulates NO and prostacyclin production, it may be useful in combating high blood pressure. More specifically, an agent of the invention may be useful
5 in therapy of three diseases of particular interest.

The first is essential hypertension, i.e. systemic high blood pressure at any location, or all around the body. For the treatment of essential hypertension, which is a systemic condition, it is preferred that a VEGF protein or agonist of the invention be delivered in a systemic manner, e.g. by systemic delivery of VEGF nucleic acids encoding VEGF
10 proteins or agonists, by gene therapy.

The second is cor pulmonale, i.e. right heart failure caused by high blood pressure in the pulmonary artery. This may be treatable by administering VEGF nucleic acids, proteins and agonists as described herein, especially by delivering VEGF DNA to the artery *via* an arterial collar (see above) with subsequent expression of the DNA to yield protein in the
15 arterial wall.

The third is primary pulmonary hypertension, i.e. high blood pressure in the lungs. This usually leads, ultimately, to heart failure and death, and is currently only treatable by means of continuous prostacyclin infusion or a lung transplant. Here, the preferred treatment technique would be to transform or transfect lung tissue with a nucleic acid
20 encoding VEGF or an agonist thereof, thereby generating VEGF *in vivo* and stimulating NO and/or prostacyclin production.

Accordingly, the invention provides the use of an agent selected from Vascular Endothelial Growth Factor (VEGF), a nucleic acid encoding VEGF, an agonist of VEGF, and a nucleic acid molecule encoding an agonist of a receptor to which VEGF binds, in
25 the manufacture of a medicament for the stimulation of nitric oxide (NO) or prostacyclin production *in vivo*.

Kits of the invention which comprise VEGF proteins, or nucleic acids, or agonists of a receptor to which VEGF binds or nucleic acids encoding such agonists are also suitable for the treatment of hypertension.

30 The invention also provides a method of treating or preventing hypertension comprising administering to a patient an effective non-toxic amount of a VEGF protein,

nucleic acid or agonist of the invention. Such treatment is effected as described herein. For primary pulmonary hypertension, the preferred method is transformation of lung tissue with a VEGF-encoding nucleic acid or a nucleic acid encoding an agonist of a receptor to which VEGF binds, as defined herein.

- 5 A further condition that can be treated according to the invention is atherosclerosis, which may be an NO-linked and/or prostacyclin-linked condition. Where the treatment of atherosclerosis is concerned, it is preferred to use a VEGF agonist as defined herein.

Thus, the invention provides the use of an agent selected from Vascular Endothelial Growth Factor (VEGF), a nucleic acid encoding VEGF, an agonist of VEGF, and a
10 nucleic acid molecule encoding an agonist of a receptor to which VEGF binds, in the manufacture of a medicament for the treatment of atherosclerosis by stimulation of NO and/or prostacyclin production *in vivo*.

The invention also provides a method of treating or preventing atherosclerosis comprising administering to a patient an effective non-toxic amount of a VEGF protein,
15 nucleic acid or agonist of the invention. Such treatment is effected as described herein.

For treatment of atherosclerosis, one preferred mode of delivery is oral delivery, e.g. in tablet form, of a VEGF protein of the invention or, preferably, of a VEGF agonist of the invention.

The following Examples illustrate the invention. The following abbreviations are
20 used:

BSA	-	bovine serum albumin
DMEM	-	Dulbecco's modified Eagle's medium
FCS	-	Fetal calf serum
HUVEC	-	Human umbilical vein endothelial cells
25 IgG	-	Immunoglobulin G
MAP kinase	-	mitogen-activated protein kinase
mAb	-	monoclonal antibody
PBS	-	phosphate buffered saline
PGI ₂	-	prostacyclin
30 cPLA ₂	-	cytosolic phospholipase A ₂
PlGF	-	placenta growth factor

SDS PAGE	-	sodium dodecyl sulphate - polyacrylamide gel electrophoresis
VEGF	-	vascular endothelial growth factor
vWF	-	von Willebrand's factor
VSMC	-	vascular smooth muscle cells

5 **Example 1**

The effect of endothelial cell (EC)-specific VEGF gene transfer on the thickening of the intima was studied using a silicone collar inserted around carotid arteries which acted both as the agent that caused intimal smooth muscle cell growth and as a reservoir for the gene and vector. The model preserved EC integrity and permitted direct extravascular
10 gene transfer without any intravascular manipulation.

Example 1.1

Gene Transfer: Intimal thickening was induced in the carotid arteries of thirty-two New Zealand White rabbits by inserting an inert silicone collar around the arteries under a general anaesthesia (Booth *et al*, Atherosclerosis (1989) 76:257-268). Gene transfer was
15 done five days after positioning of the collar by gently opening the collar under anaesthesia and injecting 500 µl plasmid/liposome complexes into the collar (i.e. on the adventitial surface of the artery). No intravascular manipulations were involved in any steps of the studies.

Plasmid/liposome Complexes: Twenty-five µg pCMV5-VEGF-164 plasmid (containing mouse VEGF cDNA (Breier *et al*, Development (1992) 114:521-32; nucleotides 1-583)
20 was complexed with 25 µl Lipofectin (BRL) while diluted to 500 µl with Ringer solution. Complexes were kept at room temperature at least 15min before the gene transfer. It was determined previously that at the concentration used in the present study plasmid/Lipofectin complexes were not toxic to rabbit aortic EC *in vitro*. Control arteries
25 were transfected with a similar plasmid/liposome complex containing *E. coli* lacZ cDNA (Kalnins *et al, supra*) (nucleotides 1-3100) expression plasmid. Plasmids used for the studies were isolated from *E. coli* cultures (DH5α) using Qiagen Mega columns and purified using three phenol/chloroform extractions and one ethanol precipitation (Ausubel
30 *et al*, eds. Current Protocols in Molecular Biology. New York, NY: Greene Publishing Associates and John Wiley & Sons (1991) 4.2.3-4.2.4) were adjusted to 1µg/µl and analysed to be free of any microbiological or endotoxin contamination (Limulus assay,

detection limit 0.2ng). Animals were sacrificed 3 (n=8), 7 (n=12) and 14 (n=12) days after the gene transfer operation; arteries were carefully removed and divided into three equal portions: the proximal third was immersion-fixed in 4% paraformaldehyde/PBS for 15 min and embedded in OCT compound (Miles Scientific) (Ylä-Herttuala *et al.*: J. Clin. Invest. 5 (1995) 95: 2692-2698). The middle third was fixed as above for 4h, rinsed in 15% sucrose for 48h and embedded in paraffin. The distal third was directly embedded in OCT compound and frozen in liquid nitrogen. In four arteries, the distal third was used for mRNA isolation and RT-PCR (see below). Ten randomly selected sections from the middle portion were used for the determination of intima/media thickness ratio (Ylä-Herttuala *et al.*: Arteriosclerosis (1986) 6; 230-236) by two independent observers without 10 knowledge of the origin of the samples. Mean values of the two independent measurements were used to calculate the results (mean \pm SD). Differences in the intima/media thickness ratios between the groups were analysed by ANOVA, followed by modified t-test (* p<0.05).

15 **RT-PCR:** Distal portions of VEGF (n=2) and lacZ (n=2) transfected arteries collected 7 days after the gene transfer were used for mRNA isolation (Micro-FastTrack, Invitrogen) and were reverse-transcribed to the first strand cDNA using AMLV reverse transcriptase (5U per reaction, Boehringer) using random hexamer primers (cDNA cycle Kit, Invitrogen) as described (Hiltunen *et al.*: Circulation (1995) 92:3297-3303). A thirty-five 20 cycle PCR was performed with Taq polymerase (Boehringer) and primers specific for the transfected pCMV5-VEGF-164 construct (5'-primer: SEQ ID No. 9; 3'-primer: SEQ ID. No. 10; PCR cycle parameters: 1min 90°C, 1min 60°C, 1min 72°C, except for the last cycle 5min). Amplified fragment with an expected length (547nt) was seen in the VEGF-transfected arteries. DNA size markers (1kb ladder, BRL) are shown on both sides of the 25 gel.

Micrographs were taken, showing the characteristics of rabbit carotid arteries 7d after VEGF or lacZ gene transfer. Immunostainings of the transfected arteries demonstrated: Control artery transfected with lacZ-plasmid/liposomes (SMC-specific MAb HHF-35, 1:500 dilution, Enzo Diagnostics) showed typical intimal thickening; Artery transfected 30 with VEGF plasmid/liposomes (SMC-specific MAb HHF-35) showed only limited intimal thickening; Endothelium was present in all studied vascular segments (serial section to A,

EC-specific MAb CD31, dilution 1:50, DAKO); No evidence of inflammation was detected in VEGF-and lacZ-transfected arteries (Macrophage-specific MAb RAM-11 dilution 1:500, DKO); Neovascularization in the adventitia of VEGF-transfected artery was seen 14 days after gene transfer (Hematoxylin-eosin staining).

- 5 The avidin-biotin horseradish peroxidase system (Vector Elite, Vector Labs) was used for the immunostainings (Ylä-Herttuala *et al*, PNAS (1990) 87: 6959-6963). Controls for the immunostainings included incubations with irrelevant class-and species-matched immunoglobulins and incubations where the primary antibodies were omitted. *In situ* hybridizations were done using an anti-sense VEGF riboprobe (583nt) synthesized from pBluescript SK plasmid (Stratagene) as described (Ylä-Herttuala *et al* (1990), *supra*). Briefly, paraffin-embedded sections were pretreated with Proteinase K, acetylated and hybridized using 35-S-UTP (DuPont, NEN)-labelled riboprobes (6×10^6 cpm/ml) at 52°C for 16h. Final wash after the hybridization was with 0.1xSSC at 60°C for 30 min. Autoradiography was used for the signal detection (Eastman-Kodak NAB-2). Control
- 10 hybridizations with a nonhybridizing sense riboprobe (Ylä-Herttuala *et al* (1990), *supra*) gave negative results. Sections were counter stained with hematoxylin.

Example 1.2

- Gene transfer was carried out as described in Example 1.1. L-NAME (70mg/kg/d) was given to the rabbits in drinking water, starting one day before VEGF (n=5) or lacZ
- 20 (n=5) gene transfer. Animals were sacrificed 7 days after gene transfer and analysed for the intima/media thickness ratio and histology as described above (Ylä-Herttuala *et al*, Arteriosclerosis (1986) 6: 230-236; Ylä-Herttuala *et al* (1990) *supra*). The difference in intimal thickening was abolished.

Example 1.3

- 25 Confluent cultures of HUVEC were washed twice with serum-free medium and incubated with this medium either in the presence of the concentrations of recombinant human VEGF indicated for 15 min, or with 10ng/ml VEGF for the times shown. In some experiments, cells were pretreated for 1h with 100µM L-NAME and subsequently treated either with or without 10ng/ml VEGF for 10 min. The medium was removed and cells
- 30 were rapidly lysed at 4°C by addition of 10mM Tris/HCl (pH 7.6), 5mM EDTA, 50mM NaCl, 30mM sodium pyrophosphate, 50mM NaF, 0.1mM Na₃VO₄, 1mM PMSF and 1%

Triton X-100 (lysis buffer). Lysates were clarified by centrifugation at 15000 xg for 10min, and immunoprecipitations were performed by incubating clarified lysates with PY20 anti-phosphotyrosine mAb for 2h at 40°C. Immunoprecipitates were collected by incubating lysates for a further 1 hour with protein A-agarose. Immunoprecipitates were washed three times with lysis buffer and proteins were then extracted with 2x SDS-PAGE sample buffer. After SDS-PAGE, immunoprecipitated proteins were transferred to membranes and then immunoblotted with PY20 mAb. This shows that VEGF induces phosphorylation of a major 205 kD protein corresponding to VEGF receptor (the major tyrosine phosphorylated proteins at 100, 125, 145, 190 and 205). L-NAME abolishes the response to VEGF.

Recombinant VEGF was added at the concentration of 25 ng/ml for times up to 2 h (nitrite production was increased from the intimal level of c. 16 μ M, and remained at 1.8-2 μ M). or at the indicated concentrations of 0, 2.5, 25 ng/ml for 10 minutes. Effect of L-NAME (100 μ M) pretreatment (1 hour) on the VEGF response was measured after addition of 25 ng/ml VEGF for 10 minutes. Nitrite production was measured using capillary detection method (Leone *et al*, in Methods in Nitric Oxide Research, Feelisch and Stanler, eds. John Wiley & Sons, New York (1996) 499-508). The level rose to c. 19 and c. 2.1 μ M in the presence of VEGF; it was reduced to the control level (c. 17 μ M) on the addition of L-NAME.

Results: It was found that, as compared with lacZ-transduced arteries, VEGF gene transfer significantly reduced intimal thickening one week after the operation (intima/media ratio 0.3 vs 1.1, <0.05, respectively). The effect was reduced after two weeks, which is probably due to the fact that the plasmid/liposome-mediated gene transfer typically only induces temporary expression of the transfected gene with maximal protein expression between 2-3 days after the gene transfer (Nabel *et al*, Ann. Rev. Physiol. (1994) 56: 741-761).

Immunohistochemical analysis of the arteries showed that intimal thickening was almost exclusively composed of SMC. Endothelial layer was present in all studied segments. No adverse effects or inflammation were detected in the transfected arteries.

Expression of the transfected VEGF was confirmed by RT-PCR using primers specific for the transgene and by *in situ* hybridization. Most of the VEGF and lacZ

expression occurred in the adventitia and outer media in fibroblasts and SMCs. Adventitial neovascularization was seen in three of the VEGF-transfected arteries 14 days after the gene transfer. No neovascularization was detected in lacZ-transfected arteries.

It has been shown previously that lacZ-plasmid/liposome gene transfer using the collar model leads to a local gene transfer in 0.05% of arterial cells. In spite of the low gene transfer efficiency, the secreted form of VEGF produced inside the collar leads to biological effects in the local arterial microenvironment, as indicated by the presence of neovascularization in three VEGF-transfected arteries 14 days after gene transfer. As in acute hypoxia, secreted VEGF is believed to reach the EC by diffusion and bind to VEGF receptors on EC.

It was hypothesized that the inhibitory effects of VEGF on intimal thickening were due to either a direct or indirect VEGF-induced EC-derived factor or activity that could, in turn, inhibit SMC proliferation. In particular, it was hypothesized that the effects of VEGF on intimal thickening were mediated through the NO pathway. This hypothesis was tested in a subset of New Zealand White rabbits (n=8) by giving the animals NO synthase inhibitor L-NAME during the gene transfer experiments. It was found that L-NAME abolished the difference in intimal thickening between VEGF-and lacZ-transfected arteries. The main target cells for VEGF in the arterial wall are EC. The only other cell types possessing VEGF receptors are monocytes but, as judged from immunocytochemistry with specific antibodies, monocytes are absent from the collared carotid arteries under these conditions.

These results (the abilities of difference in intimal thickening) were consistent with VEGF-induced inhibition of intimal thickening through the stimulation of NO production. It was therefore examined whether VEGF could directly stimulate NO production in cultures of EC. VEGF induced tyrosine phosphorylation of a major 205 kDa protein corresponding to the VEGF receptor within the concentration range 1-25ng/ml. Addition of VEGF to cultured human umbilical vein endothelial cells (HUVEC) caused a time- and concentration-dependent increase in NO production as monitored by measurement of nitrite levels. The effect of VEGF on NO production was seen as early as 30 seconds after the addition of VEGF, reached a maximum after 5 min and was sustained for up to 2 hours. The half-maximal effect of VEGF was obtained at 5ng/ml. VEGF-induced

phosphorylation and NO production were completely abolished in the presence of 100 μ M L-NAME. Thus, it is likely that VEGF gene transfer stimulates NO production in EC in the transfected arteries and limits SMC proliferation at least partially *via* an NO-mediated mechanism. The findings are compatible with the previous observations that transfection of arteries with endothelial NO synthase cDNA reduces intimal thickening (Von der Leyon
5 *et al*, PNAS (1995) 92:1137-1141).

Callow *et al* (*supra*) and Asahara *et al* (*supra*) concluded that administration of VEGF protein stimulated EC proliferation in denuded arteries, but the actual mechanisms involved in the inhibition of intimal thickening were not studied. In the collared carotid
10 artery, intimal thickening is stimulated in the presence of an anatomically intact endothelium. Therefore, it is unlikely that the inhibitory effect of arterial VEGF gene transfer on intimal thickening reported here is due to VEGF-stimulated re-endothelialization. According to these results, VEGF can directly induce NO production in HUVEC such that this is one mechanism through which VEGF can inhibit
15 intimal thickening. VEGF may also stimulate the production of other factors which can negatively regulate SMC proliferation including TGF- β or prostacyclin.

Example 2

Using plasmid/liposome complexes for gene delivery, Moloney murine leukemia virus-derived (MMLV) retroviruses, pseudotyped vesicular stomatitis virus protein-G
20 (VSV-G)-containing retroviruses and adenoviruses were delivered into the rabbit carotid artery using a silastic collar applied to the adventitia. The collar is used because 1) it provides a gene delivery reservoir; 2) no intraluminal manipulations are performed and endothelium remains anatomically intact throughout; and 3) installation of the collar induces arterial smooth muscle cell (SMC) proliferation and enhances retroviral gene
25 transfer efficiency where target cell proliferation is required.

Gene Transfer: New Zealand White rabbits (1.8-2.5kg) were used. The anaesthetic was fentanyl-fluanisone (0.3 ml/kg)/midazolam (1 mg/kg) (Ylä- Herttuala *et al*, J. Clin Invest. (1995) 95:2692-2698). A midline neck incision exposed the left carotid artery. A biologically inert 2 cm silastic collar (MediGene Oy, Kuopio, Finland) was positioned
30 around the carotid artery so that it touched the adventitia lightly at either end (Booth *et al*, *supra*). Gene transfer was performed 4-5 days after the collaring operation. For gene

transfer, animals were re-anaesthetized. The collar, which had been surgically re-exposed, was gently opened and filled with 500 µl of the gene transfer solution (see below). The incision was closed and arteries later analysed for gene transfer efficiency.

Histological Analysis: Collared arteries were carefully removed and divided into three equal parts: the proximal third was immersion-fixed in 4% paraformaldehyde/PBS (pH 7.4) for 15 min, followed by embedding into OCT compound (Miles Scientific, USA). The medial third was immersion-fixed in 4% paraformaldehyde/PBS (pH 7.4) for 4 h, rinsed in 15% sucrose (pH 7.4) for 48 h and embedded in paraffin. The distal third was embedded in OCT compound and processed for frozen sections. Ten randomly selected sections were stained with X-gal for β-galactosidase activity for 12 h and used for the determination of gene transfer efficiency (Nabel *et al*, Science (1990) 249: 1285-1288; Ylä-Herttuala *et al*, (1995) *supra*). Gene transfer efficiency was calculated as a percentage of the β-galactosidase-containing cells as a proportion of the total number of nuclei in 20 randomly selected 100X fields. Randomly selected sections from each third portion of the collared arteries were used for immunocytochemistry and analysis of cell types and/or intima/media thickness ratios (Booth *et al*, *supra*).

Cell types were identified using the following antibodies: SMC: HHF-35 mAb (1:500 dilution, Enzo Diagnostics, USA), α-actin mAb (1:1000 dilution; Sigma Chemical Co.); macrophages: RAM-11 mAb (1:1000 dilution; Dako, USA), anti-CD68 mAb (1:250 dilution; Dako); endothelial cells: anti-CD31 mAb (1:50 dilution; Dako); polymorphonuclear leukocytes: anti-CD45 mAb (1:100 dilution; Dako); and anti-rabbit T-cells: MCA 805 mAb (1:1000 dilution; Dako). The avidin-biotin-horseradish peroxidase system was used for signal detection (Vector laboratories) (Ylä-Herttuala *et al* (1995) *supra*). After immunostaining, tissue sections were counter-stained with hematoxylin.

Determination of the Proliferation Index: The proliferation index in the collared arteries was determined using the 5-bromo-2'-deoxyuridine (BrdU) labelling (Soma *et al*, Arterioscler. Thromb. (1993) 13:571-578). Briefly, New Zealand white rabbits (n=12) were injected with BrdU (40 mg/kg body weight) 3 h before sacrifice. Carotid arteries were fixed in 70% ethanol overnight and embedded in paraffin. Serial sections (20 sections per animal) were stained to detect BrdU using FITC-labelled anti-mouse IgG (Dako), following propidium iodide staining of the nuclei. The labelling index was calculated as the

percentage of the BrdU-positive nuclei. Contralateral carotid arteries were sham-operated and used as controls.

Plasmid/liposomes: pCMV- β -galactosidase (lacZ) expression plasmid (Promega) was complexed with Lipofectin reagent (BRL) as follows: 25 μ g plasmid was slowly mixed with 25 μ l Lipofectin reagent while diluted to 500 μ l with Ringer solution. No precipitates were observed in the plasmid/Lipofectin solution. The mixture was left to stand at room temperature for at least 15 min and used for gene transfer within two hours. Plasmid preparations were checked for the absence of lipopolysaccharide contamination (Limulus assay, Sigma Chemical Co.).

Retroviruses: LacZ-containing pLZRNL MMLV retroviruses (Ylä-Herttuala *et al*, (1995) *supra*; Miyahara *et al*, PNAS (1988) 85:6538-6542) or LacZ VSV-G pseudotyped retroviruses (Yee *et al*, PNAS (1994) 91:9564-9568) were used for the studies. In both, the expression of lacZ is driven by the 5' LTR. Replication-deficient LZRNL amphotrophic retroviruses were packaged in PA317 cells and used at a titer of 5×10^5 cfu/ml as described (Ylä-Herttuala *et al*, (1995) *supra*). Replication-deficient VSV-G pseudotyped retroviruses were produced in 293 GP cells using transient transfection (Yee *et al*, *supra*). Pseudotyped retroviruses were concentrated using ultracentrifugation and used at a titer of 1×10^7 cfu/ml. Before use, retroviral preparations were checked for the absence of any bacteriological contaminants or helper viruses (Yee *et al*, *supra*).

Adenoviral Vectors: Replication-deficient E1-deleted adenoviruses were used for the studies (Gosh-Choudhury *et al*, Gene (1986) 50: 161-171; Simari *et al*, J. Clin. Invest. (1996) 98: 225-235). Nuclear-targeted β -galactosidase cDNA under a β -actin-promoter and a CMV enhancer was cloned into the E1-deleted region of the adenoviral genome using homologous recombination (Gosh-Choudhury *et al*, *supra*; Simari *et al*, *supra*). Replication-deficient adenoviruses were produced in 293 cells and concentrated by ultracentrifugation. Titers of 1×10^9 pfu/ml were used for the gene transfer experiments. Adenoviral preparations were analysed for the absence of helper viruses or bacteriological contaminants (Gosh-Choudhury *et al*, *supra*).

Results: The adventitial collar led to neointimal hyperplasia 7-14 days after the operation.

The endothelium remained anatomically intact throughout the studies. BrdU labelling indicated a peak proliferation index of 23% 3 days after the operation. The neointima was

exclusively composed of SMC. Plasmid/liposome complexes led to a detectable gene transfer into the adventitia and outer media, with an efficiency of less than 0.01%. Untransfected or liposome-treated collared arteries showed no staining for β -galactosidase activity.

5 Adventitial retroviral gene transfer was less efficient without the collar, probably because retroviral gene transfer only occurs in proliferating cells. The gene transfer efficiency with replication-deficient MMLV retroviruses was low (less than 0.01%). Gene transfer efficiency with VSV-G pseudotyped retroviruses was 0.1%. Using MMLV and VSV-G retroviruses, β -galactosidase staining was observed in the adventitia and outer
10 media.

Replication-deficient adenoviruses gave efficient gene transfer, β -galactosidase staining being detected in the adventitia and outer media. Interestingly, staining was also observed in some endothelial cells and in some intimal cells. Since the lacZ adenovirus construct contained a nuclear localization signal for β -galactosidase, intense X-gal staining
15 was located in the nuclei of the transfected cells. Gene transfer efficiency was approximately 10%, as estimated from the total number of stained nuclei in the analysed sections. Some inflammatory cells were seen in VSV-G retrovirus and adenovirus-transfected arteries. No inflammatory cells were seen in the plasmid/liposome transfected arteries.

20 In this Example, the transfer of the β -galactosidase (lacZ) marker gene to the adventitia and outer media occurred with all gene transfer systems. Adenoviruses also transferred the β -galactosidase gene to some endothelial cells. After five days, adenoviral vectors produced the highest gene transfer efficiency, with up to 10% of cell showing β -galactosidase activity. Pseudotyped VSV-G retroviruses were also effective in achieving
25 gene transfer in 0.1% of cells in the adventitia and outer media. Plasmid/liposome complexes and MMLV retroviruses infected $<0.01\%$ of cells. No adverse tissue reactions were observed with any of the gene transfer systems.

Thus, replication-deficient adenoviruses, VSV-G pseudotyped retroviruses and plasmid/liposome complexes can be used for gene transfer to the arterial wall using the
30 collar method. Effects on medial SMC and even endothelium can be achieved from the adventitial side.

Example 3

In this Example, the stimulation of PGI₂ by VEGF was examined.

Cell Culture: HUVECs were obtained either from Clonetics and cultured in the manufacturer's own medium supplemented with 2% FBS, or were grown from fresh
5 umbilical cords by collagenase digestion and cultured on 1% gelatin-coated plates in medium 199 supplemented with 20% FCS and endothelial cell growth supplement (Wheeler-Jones *et al*, Biochem. J. (1996) 315:407-416). For experimental purposes, primary cultures of HUVEC were dispersed by treatment with 0.05% trypsin/0.02% EDTA for 5 min at 37°C and then replated in either 90 mm, 60 mm or 35 mm plastic
10 dishes, or onto 24-well plates. Cultures were maintained in a humidified atmosphere containing 5% CO₂ and 90% air at 37°C and used after 6-8 days or when the cells had formed a confluent monolayer.

PGI₂ Assay: Confluent cultures of HUVEC in 24-well plates were washed twice in serum-free M199 (pH 7.4) and exposed to medium containing VEGF. The PGI₂ content of cell
15 supernatants was quantified by radioimmunoassay of 6-keto-PGF_{1α}, the stable breakdown product of PGI₂ as previously described (Wheeler-Jones *et al*, *supra*).

Arachidonic Acid Release: Arachidonic acid release was determined essentially as described (Domin and Rozengurt, J. Biol. Chem. (1993) 268:8927-8934.). Confluent HUVEC cultures were incubated for 24 h with [5,6,8,9,11,12,14,15-³H]arachidonic acid
20 (1 mCi/ml, 211 Ci/mmol). The cells were then washed twice with medium 199 and incubated in 1 ml of this medium supplemented with 0.3% BSA (essential fatty-acid free) and VEGF or thrombin. After treatment, the medium was removed, centrifuged in a microcentrifuge at 16,000xg for 5 min, and the radioactivity in the supernatant was determined by counting in a scintillation counter.

Western Blotting Procedures: Treatment of quiescent cultures of cells with factors, and cell lysis were performed as described above and in the Results and Figure Legends. After SDS-PAGE, proteins were transferred to Immobilon membranes (Millipore Inc.). For MAP kinase assays, membranes were blocked using 5% non-fat dried milk in PBS, pH 7.2, and incubated for 3-5 h in PBS/0.05% Tween-20 containing primary antibody (1mg/ml)
25 as indicated. For immunoblots with antibody to cPLA₂, membranes were blocked for 3h
30 in TBST50 mM Tris/HCl, 150 mM NaCl, 0.02% (v/v) Tween 20 pH 7.4 (TBST)

containing 0.2% (w/v) I-block (Tropix), then incubated with primary antiserum in TBST. Membranes were then washed six times (10 min each wash) in TBST and incubated for 1 hour in TBST containing HRP-conjugated secondary antibody. Immunoreactive bands were visualized by chemiluminescence using HRP-conjugated anti-mouse or anti-rabbit IgG and ECLTM reagent according to the manufacturer's instructions.

MAP Kinase Assay: Cells were treated with factors as indicated, washed rapidly twice with ice-cold PBS and immediately extracted by the addition of 100ml boiling 2x SDS-PAGE sample buffer. Cell extracts were collected by scraping, heated to 95°C for 10 min and run on 12.5% acrylamide SDS-PAGE gels. Following transfer to Immobilon membranes, proteins were immunoblotted with an antibody which specifically recognizes p42 and p44 MAP kinases (Erk-1 and Erk-2) activated by phosphorylation at Tyr204 (Payne *et al*, EMBO J. (1991) 10:885-892.).

cPLA₂ Mobility Shift Assay: Confluent quiescent HUVECs in 60 mm dishes were washed twice in serum-free medium 199 (pH 7.4) and subsequently exposed, for the times indicated, to medium containing factors as detailed in the Figure legends. Cell lysates were prepared as previously described (Wheeler-Jones *et al*, *supra*). Proteins were separated by SDS-PAGE (10% acrylamide) and after transfer to membranes were immunoblotted with polyclonal anti-serum to cPLA₂ (BorschHaubold *et al*, J. Biol Chem. (1995) 270: 25885-25892; and Kramer *et al*, J. Biol. Chem. (1996) 271:27723-27729).

Assay of vWF Secretion: vWF secretion was measured by ELISA (Wheeler-Jones *et al*, *supra*) in samples of medium obtained from confluent cultures of HUVECs which had been treated with factors, as indicated. Plates were coated with the anti-vWF mAb CLBRAg35, and the lower detection limit of the assay was approximately 1.0 mU/ml.

Materials: Recombinant VEGF was obtained either from UBI or from R & D Systems. Recombinant PlGF was the gift of Professor Werner Risau and was also obtained from R & D Systems. Polyclonal cPLA₂ antiserum was kindly provided by Dr. Ruth Kramer (Eli Lilly, Indianapolis). The anti-vWF mAb CLBRAg35 was a gift from Dr. J. A. Van Mourik (Central Blood Laboratory, Amsterdam, The Netherlands). Antibody to the activated phosphorylated form of p42/p44 MAP kinase was purchased from New England Biolabs Inc. [5,6,8,9,11,12,14,15-³H]arachidonic acid, ECLTM reagents and HRP-conjugated anti-

mouse IgG were from Amersham, UK. Goat anti-rabbit HRP-conjugated IgG was obtained from Pierce Inc. All other reagents used were of the purest grade available.

Results: Confluent cultures of HUVEC were treated for various times up to 2 h with VEGF and the medium was removed at these times and assayed for the presence of 6-keto PGF_{1α}, a stable metabolic breakdown product of PGI₂. VEGF caused a time-dependent increase in the production of PGI₂ which was detectable as early as 15 min after addition of the factor, continued to increase for up to 60 min, and was sustained thereafter for up to 2 h. Control cells exhibited only a small increase in PGI₂ production during the time-course examined. VEGF stimulation of PGI₂ production was also concentration-dependent. After a 60 min incubation, an increase in PGI₂ synthesis was detectable at 5 ng/ml, was half-maximal at 10 ng/ml and reached a maximum at 15 ng/ml. It was consistently noted that VEGF-induced PGI₂ synthesis underwent a small decline at 25 ng/ml.

It was also examined whether the VEGF-related factor PlGF, a specific ligand for the Flt-1 VEGF receptor, could elicit PGI₂ synthesis in HUVEC. In 5 independent experiments, the PlGF stimulated PGI₂ synthesis significantly more weakly than VEGF. The responses of both VEGF and PlGF were also compared to that of thrombin, a potent inducer of prostanoid synthesis in endothelial cells and platelets. In several independent experiments in which the effects of VEGF, PlGF and thrombin were directly compared in parallel cultures, the mean fold increases in 6-keto PGF_{1α}, produced by 60 min incubations with 25 ng/ml VEGF, 60 ng/ml PlGF and 1 U/ml thrombin were, respectively, 2.0-, 1.3- and 4.4-fold above the mean control unstimulated level.

If VEGF stimulation of PGI₂ synthesis was mediated through activation of an isoform of PLA₂, it would be predicted that VEGF would cause a rapid mobilization of arachidonic acid from cells. To test this, HUVECs were preincubated with radio-labelled arachidonic acid for 24 h and subsequently challenged with VEGF for different times. VEGF caused an increase in label released into the medium of prelabelled cells which was evident 10 min and reached a maximum 30 min, after addition of the factor. As measured in parallel cultures, the time-course for VEGF-stimulated arachidonic acid release was very similar to that for thrombin. The concentration-dependence for VEGF-stimulated arachidonic acid release was very similar to that obtained for PGI₂ production, with a half-

maximum effect at 2.5-5 ng/ml and a maximum at 10-20 ng/ml. Similarly to the relative abilities of thrombin and VEGF to stimulate PGI₂ production, maximum VEGF-induced arachidonic acid release was consistently lower than that obtained for thrombin. In four independent experiments, VEGF and thrombin caused mean increases in the release of
5 labelled arachidonic acid of 1.6- and 3.4-fold above the basal unstimulated level, respectively. PIGF caused no detectable significant increase in arachidonic acid release from prelabelled HUVECs.

One plausible mechanism for rapid VEGF-stimulated arachidonic acid release is the direct enzymatic release of arachidonic acid catalysed by the cytosolic form of PLA₂.
10 cPLA₂ can be activated by MAP kinase-dependent phosphorylation and, similarly to the phosphorylation and activation of other enzymes including MAP kinases, the conversion of cPLA₂ to its activated form can be monitored by a shift in its mobility in SDS-PAGE gels from a fast- to a slow-migrating form. Accordingly, activation of cPLA₂ in response to VEGF was determined by western blot analysis of HUVEC extracts using a specific
15 antibody to cPLA₂ (BorschHaubold *et al*, *supra*, and Kramer *et al*, *supra*). In extracts of control unstimulated HUVECs, antibody to cPLA₂ recognized two distinct bands of approximately equal intensity and migrating with approximate Mr 97,000. Although cPLA₂ has a predicted molecular weight of 85 kDa, this protein has previously been reported to migrate in SDS-PAGE as a 97 kDa band (BorschHaubold *et al*, *supra*, and
20 Kramer *et al*, *supra*).

VEGF at 25 ng/ml caused a marked increase in the immunoreactivity of the slow-migrating form of cPLA₂ and a concomitant relative decrease in the faster-migrating form, which was detectable after 2 min, reached a maximum after 15 min, and was sustained for up to 60 min after VEGF addition. In five independent experiments, VEGF consistently
25 caused a marked increase in the immunoreactivity of the slower-migrating form of cPLA₂. The VEGF-induced decrease in the electrophoretic mobility of cPLA₂ was also concentration-dependent, with a noticeable increase in the slow-migrating form at 2.5 ng/ml and a maximum increase at 5-10 ng/ml, the highest concentration tested. While VEGF caused an apparent reduction in the level of the faster-migrating form of cPLA₂,
30 this species was evident at all VEGF concentrations and times of treatment examined. Thrombin also caused a striking shift in the mobility of cPLA₂ from a faster- to a slower-

migrating form, both of which exactly co-migrated with those detected in extracts of VEGF-treated HUVECs. Direct comparison of the effects of VEGF and thrombin in the same experiment showed that, similarly to the results obtained for PGI₂ synthesis and arachidonic acid release, thrombin caused a more marked increase in the gel retardation of cPLA₂ with virtually complete disappearance of the faster-migrating form of the enzyme. PlGF caused no detectable shift in the mobility of immunoreactive cPLA₂ from faster to slower-migrating forms.

It was further examined whether VEGF also induced vWF secretion by HUVECs. VEGF stimulated a time- and concentration-dependent production of vWF. vWF was detectable in HUVEC medium as early as 30 min after addition of 25 ng/ml VEGF to cells and continued to increase over the time-course examined reaching a level 2.5-fold above that in control cells after 3h. The concentration-dependence for the effect of VEGF was similar to that obtained for PGI₂ production with a half-maximal response at approximately 10 ng/ml and a maximum effect at 25 ng/ml, the highest concentration tested. Similarly to the results obtained for PGI₂ synthesis, the effect of VEGF was comparable to, though consistently weaker than, that of thrombin (Fig. 12A). In contrast VEGF, PlGF caused no detectable stimulation of vWF secretion in HUVEC. Comparison of the effects of VEGF and PlGF on migration of HUVECs in chemotaxis chambers showed that while VEGF stimulated a concentration-dependent increase in chemotaxis, PlGF caused no detectable increase in the concentration range 5-60 ng/ml.

VEGF activates p42/p44 MAP kinases in HUVEC. Given that MAP kinases have been implicated in cPLA₂ activation by other factors, this raised the possibility that the MAP kinase cascade might mediate VEGF-induced PGI₂ synthesis and cPLA₂ activation. The concentration-dependence for VEGF-stimulated activation of p42/p44 MAP kinases activity in confluent HUVEC was examined. Exposure of cells to VEGF for 15 min caused a detectable increase in activity at a concentration as low as 0.5 ng/ml, a half-maximal increase between 1 and 5 ng/ml, and a maximum effect at 10 ng/ml which was sustained up to 50 ng/ml. In contrast to the striking effect of VEGF, PlGF in the concentration range 1-60 ng/ml caused little, if any, detectable increase in MAP kinase activity in HUVEC. VEGF stimulation of p42/p44 MAP kinases was completely inhibited by a 30 min pretreatment with PD98059, a selective inhibitor of MAP kinase kinase (Dudley *et al*,

PNAS USA (1995) 92:7686-7689), the dual-specificity threonine and tyrosine kinase which specifically phosphorylates and activates p42/p44 MAP kinases. The effect of the inhibitor was concentration-dependent with greater than 50% inhibition at 5 mM and a reduction of MAP kinase activity to the control, unstimulated level at 10-20 mM.

5 The role of the MAP kinase cascade in the VEGF-induced arachidonic acid mobilization pathway was initially investigated by examining the effect of PD98059 on PGI₂ synthesis. Pretreatment of HUVEC with 30 mM PD98059 for 30 min completely inhibited PGI₂ production induced by a subsequent 60 min incubation with either 25 ng/ml VEGF or 1 U/ml thrombin. The effect of PD98059 on VEGF-induced PGI₂ synthesis was
10 concentration-dependent with a half-maximal effect at approximately 5 mM and a maximum inhibitory effect on stimulated PGI₂ production at 10 mM. It was also noted that PD98059 reduced PGI₂ production in VEGF-treated cells to below the values measured in control cells, though this effect was only apparent at concentrations of the inhibitor greater than 10 mM..

15 It was next tested whether PD98059 had any effect on the VEGF-induced decrease in the electrophoretic mobility of cPLA₂. Pretreatment with PD98059 at 25 mM completely blocked the VEGF-stimulated increase in the slow-migrating form of cPLA₂. Similarly to the effects of PD98059 on PGI₂ synthesis, the inhibitor not only reversed the VEGF-dependent decrease in cPLA₂ gel mobility, but also decreased the immunoreactivity
20 of the slower-migrating form below, and increased that of the faster-migrating form above control levels.

 The selectivity of the effect of PD98059 for VEGF-induced cPLA₂ activation and PGI₂ synthesis was investigated by testing whether VEGF-stimulated vWF production was also susceptible to inhibition by PD98059. In contrast to the effect of PD98059 on cPLA₂
25 activation and PGI₂ synthesis, pretreatment of HUVECs with the MAP kinase kinase inhibitor at 25 mM neither prevented nor significantly reduced the secretion of vWF caused by a subsequent addition of VEGF. PD98059 also had no effect either on basal or thrombin-stimulated vWF secretion.

Summary: VEGF stimulated a time- and concentration-dependent increase in PGI₂
30 synthesis which was detectable within 15 min, half-maximal at 10 ng/ml and maximal at 15 ng/ml after 60 min. In 10 independent experiments, mean maximum VEGF-stimulated

PGI₂ synthesis was 2-fold above basal levels at 25 ng/ml after 60 min. The VEGF-related factor, placenta growth factor (PIGF), induced a much weaker 1.3-fold increase. and thrombin (1 U/ml, 60 min) induced a mean maximum increase of 4.4-fold above control levels. VEGF stimulated the release of arachidonic acid from HUVEC with a similar concentration-dependence to that obtained for PGI₂ synthesis but with more rapid kinetics: half-maximum arachidonic acid mobilization occurred after 10 min and was maximal after 30 min. Measurement of cytosolic phospholipase A₂ (cPLA₂) activation using mobility shift in SDS-PAGE as a marker of activation, showed that VEGF stimulated cPLA₂ activity in a time- and concentration-dependent manner: an increase in the slow-migrating and activated form of cPLA₂ occurred as early as 2 min and was detectable as low as 2.5 ng/ml, and reached a maximum after 15 min and at 5 ng/ml. Similar to other agents which induce PGI₂ synthesis, VEGF also caused a striking time- and concentration-dependent increase in secretion of von Willebrand factor (vWF) in HUVEC which was detectable within 30 min, half-maximal at 10 ng/ml and reached 2.5-fold above control levels after a 3 h treatment with 25 ng/ml VEGF. VEGF induced a rapid and transient activation of p42/p44 MAP kinases which was detectable as low as 1 ng/ml and reached a maximum at 5-10 ng/ml. In contrast, PIGF had little effect on MAP kinase activity. PD98059, a selective inhibitor of MAP kinase kinase, caused complete inhibition of VEGF-stimulated MAP kinase activity, PGI₂ synthesis and cPLA₂ gel retardation, but had no effect on VEGF-induced vWF secretion. These findings provide the first evidence that VEGF can stimulate PGI₂ synthesis *via* cPLA₂-mediated arachidonic acid release. These findings also indicate that VEGF stimulation of this biosynthetic pathway may occur, at least in part, *via* activation of p42/p44 MAP kinases.

The results presented here show that VEGF stimulates a striking time- and concentration-dependent increase in PGI₂ production in HUVEC. The effect of VEGF was weaker than that of thrombin, though the responses to the two factors varied on average by a factor of only 2.5. VEGF stimulates the rapid mobilization of arachidonic acid from human endothelial cells and, as judged by a gel retardation assay, the rapid activation of cPLA₂. The method used for determining the effect of VEGF on arachidonic acid mobilization is based upon the release of ³H-label from cells prelabelled with ³H-arachidonic acid. Since the studies of arachidonic acid release were performed with BSA

present in the medium, it is highly likely that the major product released from HUVECs is ^3H -arachidonic acid. The concentration-dependencies for VEGF-induced PGI_2 production, arachidonic acid release and cPLA_2 activation were similar to each other (all within the range 5-10 ng/ml). These results indicate that VEGF stimulation of arachidonic acid release and PGI_2 synthesis are mediated through high-affinity receptors for this factor in HUVECs.

Arachidonic acid release and the cPLA_2 mobility shift both occurred more rapidly than PGI_2 synthesis, consistent with the notion that increased production of PGI_2 is very likely, at least in part, to be a direct consequence of increased cPLA_2 activity and a subsequent increase in the availability of intracellular arachidonic acid, the substrate for the constitutive enzyme COX-1. Since VEGF is known to stimulate phospholipase C-g tyrosine phosphorylation and phosphoinositide metabolism, it is entirely plausible that other mechanisms including the sequential action of phospholipase C-g (and/or phospholipase D) and diacyl- and monacylglycerol lipases, may also contribute to VEGF-induced arachidonic acid mobilization of PGI_2 synthesis.

It was also investigated whether the VEGF-related factor, PlGF, a specific ligand for Flt-1, could stimulate the generation of PGI_2 . The results suggest that PlGF can induce PGI_2 synthesis but more weakly than VEGF. No significant effect of PlGF was detected on either arachidonic acid release or cPLA_2 mobility shift. Since PlGF binds with high-affinity only to the Flt-1 VEGF receptor, these results are most consistent with the conclusion that stimulation of PGI_2 production by VEGF in HUVECs is mediated primarily by KDR/Flk-1 receptors, but that Flt-1 may also contribute to stimulation of this pathway. However, any Flt-1-mediated PGI_2 synthesis would appear to involve a mechanism other than cPLA_2 phosphorylation. At least two explanations could account for the relatively lower response to PlGF. Either Flt-1 induces a weaker response, and/or Flt-1 is present at lower levels than KDR/Flk-1.

The results presented herein show that VEGF induced secretion of vWF in a time- and concentration-dependent manner. The concentration-dependence for the effect of VEGF on vWF secretion agreed closely with that for PGI_2 synthesis and arachidonic acid mobilization. In contrast to the relatively weak effect of PlGF on PGI_2 synthesis, PlGF did not stimulate a detectable increase in vWF release by HUVECs.

The finding that VEGF causes a rapid shift in the electrophoretic mobility of cPLA₂ is consistent with increased phosphorylation, and hence activation, of the p42/p44 MAP kinases. This is supported by the finding presented here that thrombin, which is known to stimulate cPLA₂ phosphorylation and activation in platelets, caused a similar shift in cPLA₂ mobility in HUVECs. The results presented here also show that VEGF activates p42/p44 MAP kinases and that inhibition of this pathway with a selective inhibitor of MAP kinase kinase, PD98059, blocks PGI₂ production and the increased gel retardation of cPLA₂ induced by VEGF. The effect of PD98059 was at least partially selective since it had no effect on vWF secretion stimulated either by VEGF or by thrombin.

In contrast to the striking effect of VEGF, PlGF failed to significantly increase MAP kinase activity. The apparent inability of PlGF to stimulate MAP kinase activity is broadly in accord with the much weaker effect of this factor on PGI₂ production compared to VEGF and with the apparent inability of this factor to promote cPLA₂ gel retardation.

Assays of PGI₂ production in HUVEC indicated the presence of a significant basal level of synthesis. Interestingly the MAP kinase kinase inhibitor also abolished PGI₂ production in control, unstimulated cells. Consistent with the mediation of PGI₂ generation through activation of the MAP kinase cascade, PD98059 also reduced the level of the slow-migrating phosphorylated form of cPLA₂ below the control level and increased that of the fast-migrating less phosphorylated form above the control level. Assays of MAP kinase activity suggested a significant level of basal activity which was also inhibitable by PD98059. These findings suggest that MAP kinase activation may be responsible not only for VEGF- and thrombin-dependent cPLA₂ activation, but that basal MAP kinase activity may also be required for the maintenance of constitutive cPLA₂ activity and PGI₂ production.

It is plausible that other signalling mechanisms may contribute to VEGF stimulation of PGI₂ synthesis and arachidonic acid release, including elevation of intracellular [Ca²⁺].

VEGF has to date primarily been regarded as an angiogenic factor which promotes endothelial cell growth and migration. The findings presented here reveal additional activities, and may therefore have important implications both for the regulation of endothelial function and for understanding the function of VEGF, as well as for the

treatment and prevention of blood vessel disorders such as stenosis, restenosis, atherosclerosis and hypertension.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT:

- (A) NAME: EUROGENE LIMITED
- (B) STREET: Marquis House, 67/68 Jermyn Street
- (C) CITY: London
- (D) STATE: N/A
- (E) COUNTRY: United Kingdom
- (F) POSTAL CODE (ZIP): SW1Y 6NY

(ii) TITLE OF INVENTION: THERAPEUTIC USE OF GROWTH FACTOR, AND
DELIVERY DEVICE, ESPECIALLY FOR THE TREATMENT OF INTIMAL
HYPERPLASIA

(iii) NUMBER OF SEQUENCES: 10

(iv) COMPUTER READABLE FORM:

- (A) MEDIUM TYPE: Floppy disk
- (B) COMPUTER: IBM PC compatible
- (C) OPERATING SYSTEM: PC-DOS/MS-DOS
- (D) SOFTWARE: PatentIn Release #1.0, Version #1.30 (EPO)

(v) CURRENT APPLICATION DATA:

APPLICATION NUMBER: WO (not yet known)

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 441 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..441

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

ATG AAC TTT CTG CTG TCT TGG GTG CAT TGG AGC CTC GCC TTG CTG CTC	48
Met Asn Phe Leu Leu Ser Trp Val His Trp Ser Leu Ala Leu Leu Leu	
1 5 10 15	
TAC CTC CAC CAT GCC AAG TGG TCC CAG GCT GCA CCC ATG GCA GAA GGA	96
Tyr Leu His His Ala Lys Trp Ser Gln Ala Ala Pro Met Ala Glu Gly	
20 25 30	
GGA GGG CAG AAT CAT CAC GAA GTG GTG AAG TTC ATG GAT GTC TAT CAG	144
Gly Gly Gln Asn His His Glu Val Val Lys Phe Met Asp Val Tyr Gln	
35 40 45	
CGC AGC TAC TGC CAT CCA ATC GAG ACC CTG GTG GAC ATC TTC CAG GAG	192
Arg Ser Tyr Cys His Pro Ile Glu Thr Leu Val Asp Ile Phe Gln Glu	
50 55 60	
TAC CCT GAT GAG ATC GAG TAC ATC TTC AAG CCA TCC TGT GTG CCG CTG	240
Tyr Pro Asp Glu Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu	
65 70 75 80	
ATG CGA TGC GGG GGC TGC TGC AAT GAC GAG GGC CTG GAG TGT GTG CCC	288
Met Arg Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro	
85 90 95	
ACT GAG GAG TCC AAC ATC ACC ATG CAG ATT ATG CGG ATC AAA CCT CAC	336
Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys Pro His	
100 105 110	
CAA GGC CAG CAC ATA GGA GAG ATG AGC TTC CTA CAG CAC AAC AAA TGT	384
Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln His Asn Lys Cys	
115 120 125	
GAA TGC AGA CCA AAG AAA GAT AGA GCA AGA CAA GAA AAA TGT GAC AAG	432
Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg Gln Glu Lys Cys Asp Lys	
130 135 140	
CCG AGG CGG	441
Pro Arg Arg	
145	

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 147 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

Met Asn Phe Leu Leu Ser Trp Val His Trp Ser Leu Ala Leu Leu Leu
 1 5 10 15
 Tyr Leu His His Ala Lys Trp Ser Gln Ala Ala Pro Met Ala Glu Gly
 20 25 30
 Gly Gly Gln Asn His His Glu Val Val Lys Phe Met Asp Val Tyr Gln
 35 40 45
 Arg Ser Tyr Cys His Pro Ile Glu Thr Leu Val Asp Ile Phe Gln Glu
 50 55 60
 Tyr Pro Asp Glu Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu
 65 70 75 80
 Met Arg Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro
 85 90 95
 Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys Pro His
 100 105 110
 Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln His Asn Lys Cys
 115 120 125
 Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg Gln Glu Lys Cys Asp Lys
 130 135 140
 Pro Arg Arg
 145

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 573 base pairs

00446260

(B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

(A) NAME/KEY: CDS
 (B) LOCATION:1..573

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

ATG AAC TTT CTG CTG TCT TGG GTG CAT TGG AGC CTC GCC TTG CTG CTC	48
Met Asn Phe Leu Leu Ser Trp Val His Trp Ser Leu Ala Leu Leu Leu	
150 155 160	
TAC CTC CAC CAT GCC AAG TGG TCC CAG GCT GCA CCC ATG GCA GAA GGA	96
Tyr Leu His His Ala Lys Trp Ser Gln Ala Ala Pro Met Ala Glu Gly	
165 170 175	
GGA GGG CAG AAT CAT CAC GAA GTG GTG AAG TTC ATG GAT GTC TAT CAG	144
Gly Gly Gln Asn His His Glu Val Val Lys Phe Met Asp Val Tyr Gln	
180 185 190 195	
CGC AGC TAC TGC CAT CCA ATC GAG ACC CTG GTG GAC ATC TTC CAG GAG	192
Arg Ser Tyr Cys His Pro Ile Glu Thr Leu Val Asp Ile Phe Gln Glu	
200 205 210	
TAC CCT GAT GAG ATC GAG TAC ATC TTC AAG CCA TCC TGT GTG CCG CTG	240
Tyr Pro Asp Glu Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu	
215 220 225	
ATG CGA TGC GGG GGC TGC TGC AAT GAC GAG GGC CTG GAG TGT GTG CCC	288
Met Arg Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro	
230 235 240	
ACT GAG GAG TCC AAC ATC ACC ATG CAG ATT ATG CGG ATC AAA CCT CAC	336
Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys Pro His	
245 250 255	

564T30" 35446250

56

CAA GGC CAG CAC ATA GGA GAG ATG AGC TTC CTA CAG CAC AAC AAA TGT 384
 Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln His Asn Lys Cys
 260 265 270 275

GAA TGC AGA CCA AAG AAA GAT AGA GCA AGA CAA GAA AAA CCC TGT GGG 432
 Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg Gln Glu Lys Pro Cys Gly
 280 285 290

CCT TGC TCA GAG CGG AGA AAG CAT TTG TTT GTA CAA GAT CCG CAG ACG 480
 Pro Cys Ser Glu Arg Arg Lys His Leu Phe Val Gln Asp Pro Gln Thr
 295 300 305

TGT AAA TGT TCC TGC AAA AAC ACA GAC TCG CGT TGC AAG GCG AGG CAG 528
 Cys Lys Cys Ser Cys Lys Asn Thr Asp Ser Arg Cys Lys Ala Arg Gln
 310 315 320

CTT GAG TTA AAC GAA CGT ACT TGC AGA TGT GAC AAG CCG AGG CGG 573
 Leu Glu Leu Asn Glu Arg Thr Cys Arg Cys Asp Lys Pro Arg Arg
 325 330 335

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 191 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

Met Asn Phe Leu Leu Ser Trp Val His Trp Ser Leu Ala Leu Leu Leu
 1 5 10 15

Tyr Leu His His Ala Lys Trp Ser Gln Ala Ala Pro Met Ala Glu Gly
 20 25 30

Gly Gly Gln Asn His His Glu Val Val Lys Phe Met Asp Val Tyr Gln
 35 40 45

Arg Ser Tyr Cys His Pro Ile Glu Thr Leu Val Asp Ile Phe Gln Glu
 50 55 60

Tyr Pro Asp Glu Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu
 65 70 75 80

654743 "3346260

57

Met Arg Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro
85 90 95

Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys Pro His
100 105 110

Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln His Asn Lys Cys
115 120 125

Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg Gln Glu Lys Pro Cys Gly
130 135 140

Pro Cys Ser Glu Arg Arg Lys His Leu Phe Val Gln Asp Pro Gln Thr
145 150 155 160

Cys Lys Cys Ser Cys Lys Asn Thr Asp Ser Arg Cys Lys Ala Arg Gln
165 170 175

Leu Glu Leu Asn Glu Arg Thr Cys Arg Cys Asp Lys Pro Arg Arg
180 185 190

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 645 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..645

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

ATG AAC TTT CTG CTG TCT TGG GTG CAT TGG AGC CTC GCC TTG CTG CTC

58

Met Asn Phe Leu Leu Ser Trp Val His Trp Ser Leu Ala Leu Leu Leu	
195 200 205	
TAC CTC CAC CAT GCC AAG TGG TCC CAG GCT GCA CCC ATG GCA GAA GGA	96
Tyr Leu His His Ala Lys Trp Ser Gln Ala Ala Pro Met Ala Glu Gly	
210 215 220	
GGA GGG CAG AAT CAT CAC GAA GTG GTG AAG TTC ATG GAT GTC TAT CAG	144
Gly Gly Gln Asn His His Glu Val Val Lys Phe Met Asp Val Tyr Gln	
225 230 235	
CGC AGC TAC TGC CAT CCA ATC GAG ACC CTG GTG GAC ATC TTC CAG GAG	192
Arg Ser Tyr Cys His Pro Ile Glu Thr Leu Val Asp Ile Phe Gln Glu	
240 245 250 255	
TAC CCT GAT GAG ATC GAG TAC ATC TTC AAG CCA TCC TGT GTG CCG CTG	240
Tyr Pro Asp Glu Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu	
260 265 270	
ATG CGA TGC GGG GGC TGC TGC AAT GAC GAG GGC CTG GAG TGT GTG CCC	288
Met Arg Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro	
275 280 285	
ACT GAG GAG TCC AAC ATC ACC ATG CAG ATT ATG CGG ATC AAA CCT CAC	336
Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys Pro His	
290 295 300	
CAA GGC CAG CAC ATA GGA GAG ATG AGC TTC CTA CAG CAC AAC AAA TGT	384
Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln His Asn Lys Cys	
305 310 315	
GAA TGC AGA CCA AAG AAA GAT AGA GCA AGA CAA GAA AAA AAA TCA GTT	432
Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg Gln Glu Lys Lys Ser Val	
320 325 330 335	
CGA GGA AAG GGA AAG GGG CAA AAA CGA AAG CGC AAG AAA TCC CGG TAT	480
Arg Gly Lys Gly Lys Gly Gln Lys Arg Lys Arg Lys Lys Ser Arg Tyr	
340 345 350	
AAG TCC TGG AGC GTG CCC TGT GGG CCT TGC TCA GAG CGG AGA AAG CAT	528
Lys Ser Trp Ser Val Pro Cys Gly Pro Cys Ser Glu Arg Arg Lys His	
355 360 365	
TTG TTT GTA CAA GAT CCG CAG ACG TGT AAA TGT TCC TGC AAA AAC ACA	576

664733-0346260

59

Leu Phe Val Gln Asp Pro Gln Thr Cys Lys Cys Ser Cys Lys Asn Thr
 370 375 380

GAC TCG CGT TGC AAG GCG AGG CAG CTT GAG TTA AAC GAA CGT ACT TGC 624
 Asp Ser Arg Cys Lys Ala Arg Gln Leu Glu Leu Asn Glu Arg Thr Cys
 385 390 395

AGA TGT GAC AAG CCG AGG CCG 645
 Arg Cys Asp Lys Pro Arg Arg
 400 405

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 215 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

Met Asn Phe Leu Leu Ser Trp Val His Trp Ser Leu Ala Leu Leu Leu
 1 5 10 15

Tyr Leu His His Ala Lys Trp Ser Gln Ala Ala Pro Met Ala Glu Gly
 20 25 30

Gly Gly Gln Asn His His Glu Val Val Lys Phe Met Asp Val Tyr Gln
 35 40 45

Arg Ser Tyr Cys His Pro Ile Glu Thr Leu Val Asp Ile Phe Gln Glu
 50 55 60

Tyr Pro Asp Glu Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu
 65 70 75 80

Met Arg Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro
 85 90 95

Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys Pro His
 100 105 110

Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln His Asn Lys Cys
 115 120 125

66446260

60

Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg Gln Glu Lys Lys Ser Val
 130 135 140

Arg Gly Lys Gly Lys Gly Gln Lys Arg Lys Arg Lys Lys Ser Arg Tyr
 145 150 155 160

Lys Ser Trp Ser Val Pro Cys Gly Pro Cys Ser Glu Arg Arg Lys His
 165 170 175

Leu Phe Val Gln Asp Pro Gln Thr Cys Lys Cys Ser Cys Lys Asn Thr
 180 185 190

Asp Ser Arg Cys Lys Ala Arg Gln Leu Glu Leu Asn Glu Arg Thr Cys
 195 200 205

Arg Cys Asp Lys Pro Arg Arg
 210 215

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 696 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(ix) FEATURE:

- (A) NAME/KEY: CDS
- (B) LOCATION: 1..696

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

ATG AAC TTT CTG CTG TCT TGG GTG CAT TGG AGC CTC GCC TTG CTG CTC
 Met Asn Phe Leu Leu Ser Trp Val His Trp Ser Leu Ala Leu Leu Leu
 220 225 230

48

604T30" 3B426260

61

TAC CTC CAC CAT GCC AAG TGG TCC CAG GCT GCA CCC ATG GCA GAA GGA	96
Tyr Leu His His Ala Lys Trp Ser Gln Ala Ala Pro Met Ala Glu Gly	
235 240 245	
GGA GGG CAG AAT CAT CAC GAA GTG GTG AAG TTC ATG GAT GTC TAT CAG	144
Gly Gly Gln Asn His His Glu Val Val Lys Phe Met Asp Val Tyr Gln	
250 255 260	
CGC AGC TAC TGC CAT CCA ATC GAG ACC CTG GTG GAC ATC TTC CAG GAG	192
Arg Ser Tyr Cys His Pro Ile Glu Thr Leu Val Asp Ile Phe Gln Glu	
265 270 275	
TAC CCT GAT GAG ATC GAG TAC ATC TTC AAG CCA TCC TGT GTG CCG CTG	240
Tyr Pro Asp Glu Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu	
280 285 290 295	
ATG CGA TGC GGG GGC TGC TGC AAT GAC GAG GGC CTG GAG TGT GTG CCC	288
Met Arg Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro	
300 305 310	
ACT GAG GAG TCC AAC ATC ACC ATG CAG ATT ATG CGG ATC AAA CCT CAC	336
Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys Pro His	
315 320 325	
CAA GGC CAG CAC ATA GGA GAG ATG AGC TTC CTA CAG CAC AAC AAA TGT	384
Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln His Asn Lys Cys	
330 335 340	
GAA TGC AGA CCA AAG AAA GAT AGA GCA AGA CAA GAA AAA AAA TCA GTT	432
Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg Gln Glu Lys Lys Ser Val	
345 350 355	
CGA GGA AAG GGA AAG GGG CAA AAA CGA AAG CGC AAG AAA TCC CGG TAT	480
Arg Gly Lys Gly Lys Gly Gln Lys Arg Lys Arg Lys Lys Ser Arg Tyr	
360 365 370 375	
AAG TCC TGG AGC GTG TAC GTT GGT GCC CGC TGC TGT CTA ATG CCC TGG	528
Lys Ser Trp Ser Val Tyr Val Gly Ala Arg Cys Cys Leu Met Pro Trp	
380 385 390	
AGC CTC CCT GGC CCC CAT CCC TGT GGG CCT TGC TCA GAG CGG AGA AAG	576
Ser Leu Pro Gly Pro His Pro Cys Gly Pro Cys Ser Glu Arg Arg Lys	
395 400 405	

664490" 33426260

CAT TTG TTT GTA CAA GAT CCG CAG ACG TGT AAA TGT TCC TGC AAA AAC 624
 His Leu Phe Val Gln Asp Pro Gln Thr Cys Lys Cys Ser Cys Lys Asn
 410 415 420

ACA GAC TCG CGT TGC AAG GCG AGG CAG CTT GAG TTA AAC GAA CGT ACT 672
 Thr Asp Ser Arg Cys Lys Ala Arg Gln Leu Glu Leu Asn Glu Arg Thr
 425 430 435

TGC AGA TGT GAC AAG CCG AGG CGG 696
 Cys Arg Cys Asp Lys Pro Arg Arg
 440 445

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 232 amino acids
- (B) TYPE: amino acid
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

Met Asn Phe Leu Leu Ser Trp Val His Trp Ser Leu Ala Leu Leu Leu
 1 5 10 15

Tyr Leu His His Ala Lys Trp Ser Gln Ala Ala Pro Met Ala Glu Gly
 20 25 30

Gly Gly Gln Asn His His Glu Val Val Lys Phe Met Asp Val Tyr Gln
 35 40 45

Arg Ser Tyr Cys His Pro Ile Glu Thr Leu Val Asp Ile Phe Gln Glu
 50 55 60

Tyr Pro Asp Glu Ile Glu Tyr Ile Phe Lys Pro Ser Cys Val Pro Leu
 65 70 75 80

Met Arg Cys Gly Gly Cys Cys Asn Asp Glu Gly Leu Glu Cys Val Pro
 85 90 95

Thr Glu Glu Ser Asn Ile Thr Met Gln Ile Met Arg Ile Lys Pro His
 100 105 110

664730" 8476250

63

Gln Gly Gln His Ile Gly Glu Met Ser Phe Leu Gln His Asn Lys Cys
 115 120 125

Glu Cys Arg Pro Lys Lys Asp Arg Ala Arg Gln Glu Lys Lys Ser Val
 130 135 140

Arg Gly Lys Gly Lys Gly Gln Lys Arg Lys Arg Lys Lys Ser Arg Tyr
 145 150 155 160

Lys Ser Trp Ser Val Tyr Val Gly Ala Arg Cys Cys Leu Met Pro Trp
 165 170 175

Ser Leu Pro Gly Pro His Pro Cys Gly Pro Cys Ser Glu Arg Arg Lys
 180 185 190

His Leu Phe Val Gln Asp Pro Gln Thr Cys Lys Cys Ser Cys Lys Asn
 195 200 205

Thr Asp Ser Arg Cys Lys Ala Arg Gln Leu Glu Leu Asn Glu Arg Thr
 210 215 220

Cys Arg Cys Asp Lys Pro Arg Arg
 225 230

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

TCGATCCATG AACTTTCTGC

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20 base pairs
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

TCCGTTTAAC TCAAGCTGCC

CLAIMS

1. Use of an agent that is an agonist of a receptor to which VEGF binds, or a nucleic acid encoding the agonist, for the manufacture of a medicament for the treatment or prevention of intimal hyperplasia of a blood vessel, where the endothelium is wholly or largely intact.
2. Use according to claim 1, wherein the blood vessel is an artery.
3. Use according to claim 1 or claim 2, for the treatment or prevention of stenosis induced by a surgical procedure or associated with pulmonary artery hypertension.
4. Use according to claim 3, wherein the surgical procedure is angioplasty, coronary bypass surgery, surgical anastomosis or endarterectomy.
5. Use according to any preceding claim, for the treatment or prevention of stenosis of the blood vessel.
6. Use according to any preceding claim, for the treatment or prevention of restenosis of the blood vessel.
7. Use according to any preceding claim, wherein the agent is a protein having the function of human VEGF, or a nucleic acid encoding the protein.
8. Use according to claim 7, wherein the protein has the sequence of SEQ. ID No. 2, 4, 6 or 8, or an active fragment thereof.
9. Use according to any of claims 6 to 8, wherein the agent is a nucleic acid in association with a viral or non-viral vector.
10. An implant for therapeutic use, adapted to be placed at or near the site of hyperplasia to be treated or prevented, and containing an agent as defined in any preceding claim.
11. An implant according to claim 10, which is a silastic implant or a biodegradable implant.
12. An implant according to claim 10 or 11, which is in the form of a collar for fitting around a blood vessel at or near the site of the hyperplasia to be treated or prevented.
13. An implant according to any of claims 10 to 12, having an outer wall substantially impermeable to the agent comprised in it.

14. Use of an agent as defined in any of claims 6 to 9, for the manufacture of a medicament for therapy of a condition that can be treated or prevented by stimulation of nitric oxide (NO) and/or prostacyclin production *in vivo*.
15. Use according to claim 14, wherein the condition is hypertension, e.g. essential hypertension, primary pulmonary hypertension or cor pulmonale.
16. A device for use in the delivery of a therapeutic agent to a blood vessel in a patient, which comprises a body adapted to provide a seal around the vessel, the agent being held within or associated with the device so that, in use, the agent comes into contact with the adventitial surface of the vessel.
17. A device according to claim 16, which defines a reservoir between the body wall and the vessel's adventitial surface, the reservoir being at least part-filled by a pharmaceutical formulation containing the agent to be delivered.
18. A device according to claim 17, wherein the formulation is in the form of a fluid or gel that is injectable into the reservoir or a paste.
19. A device according to claim 18, wherein the material of the body portion is self-sealing.
20. A device according to any of claims 17 to 19, wherein the reservoir can contain up to 10 ml of fluid, preferably at least 2-5 ml.
21. A device according to any of claims 17 to 20, wherein the thickness of the body material is generally constant along its length, the reservoir being formed in use by a ballooning of the first body portion between spaced-apart portions that seal against the vessel.
22. A device according to any of claims 17 to 20, wherein the thickness of the body material is smaller in an intermediate portion than at spaced-apart sealing portions that seal against the vessel, the reduced thickness forming the reservoir.
23. A device according to any of claims 16 to 22, wherein the inner surface of the body comprises a sponge-like material which is capable of being impregnated with a pharmaceutical formulation containing the agent.
24. A device according to any of claims 16 to 22, wherein the inner surface of the body is impregnated with a pharmaceutical formulation containing the agent.

25. A device according to any of claims 16 to 24, wherein the material of the body is biodegradable.
26. A device according to claims 25, wherein said material is gelatin, alginate or collagen.
- 5 27. A device according to any of claims 16 to 26, wherein the body is moulded or extruded.
28. A device according to any of claims 16 to 27, wherein the body comprises flexible seal portions that can accommodate expansion of the blood vessel caused by pulsatile blood flow.
- 10 29. A device according to any of claims 16 to 28, wherein the body comprises elongate seal portions 8-15 mm long.
30. A device according to any of claims 16 to 29, wherein the body is generally tubular.
31. A device according to any of claims 16 to 29, wherein the body has two generally
- 15 tubular portions which are branched, to form a generally Y- or T-shaped body.
32. A device according to any one of claims 16 to 29, wherein the body has three generally tubular portions which, at least in use, are branched, to form a generally X-shaped body.
33. A device according to claim 31 or claim 32, wherein one body portion is generally
- 20 arcuate in cross-section transverse to its longitudinal extent so as to enable it to surround the exposed portion of a first blood vessel when that vessel is part-embedded in tissue, and longitudinally-extending edges of the first body portion are arranged to be sealed, in use, to the adventitial wall of the first blood vessel or to adjacent tissue.
34. A device according to any of claims 16 to 33, wherein the body has a longitudinal
- 25 slit, to facilitate its fitment over the blood vessel.
35. A device according to any of claims 16 to 34, wherein the body includes an inner layer or helical reinforcement, to increase torsional strength.
36. A method for delivering an agent as defined in any of claims 1 to 10 to a blood
- 30 vessel, which comprises placing a device according to any of claims 16 to 35 around the vessel and, if not already present, introducing the agent into the device.

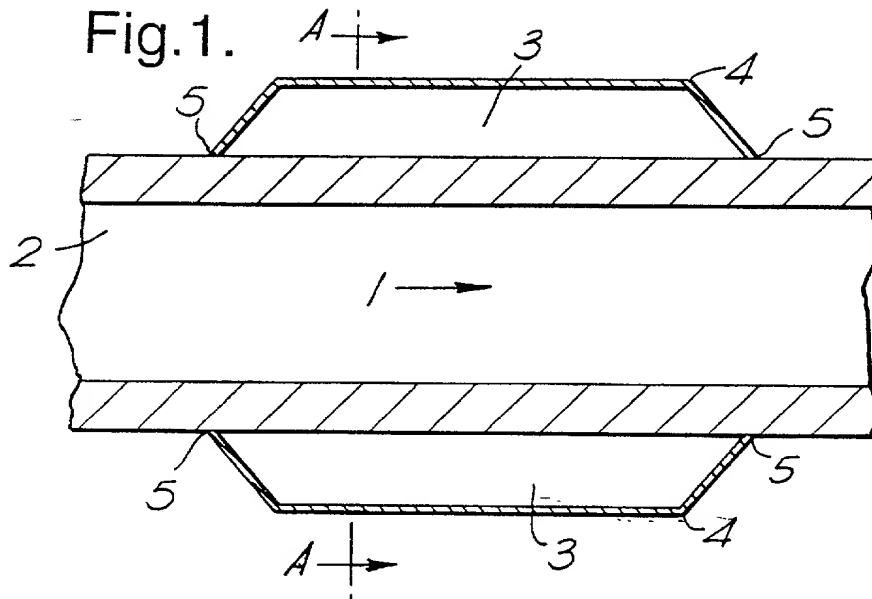


Fig.2.

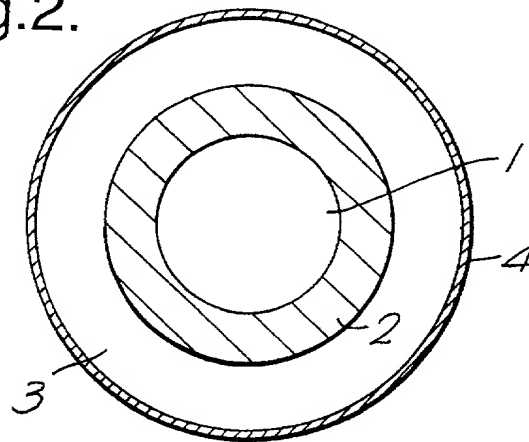


Fig.3.

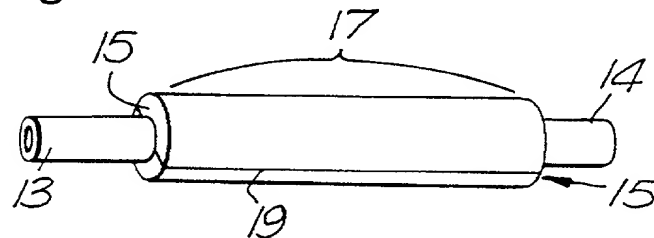


Fig.4.

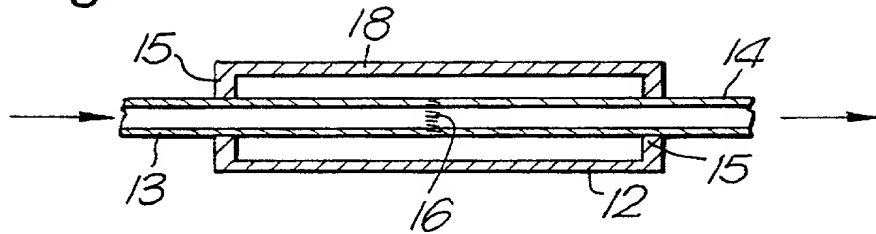


Fig.5.

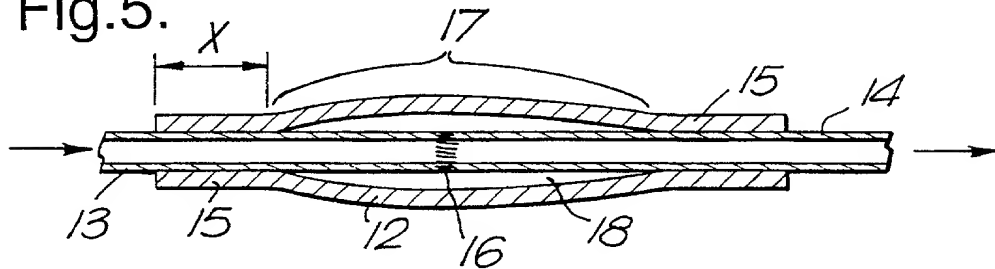


Fig.6.

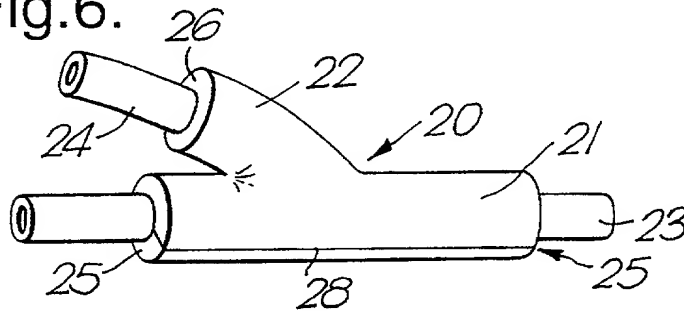


Fig.7.

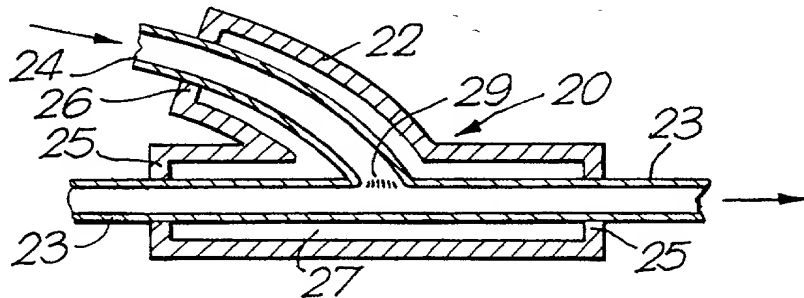
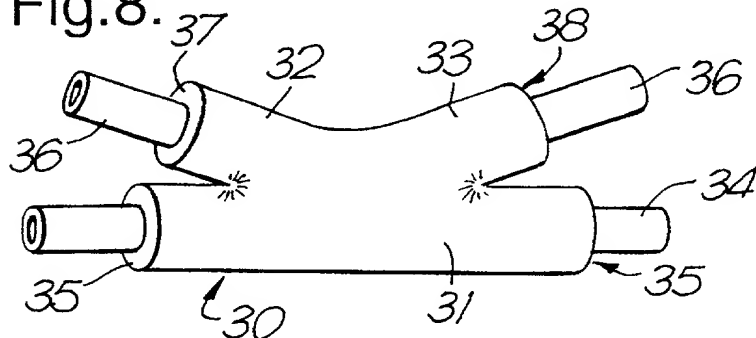
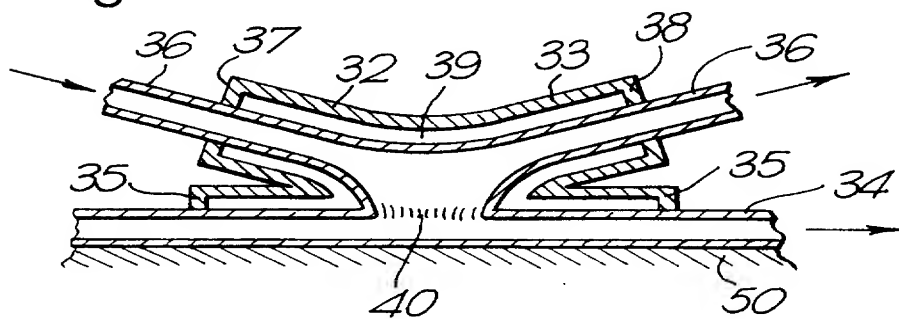
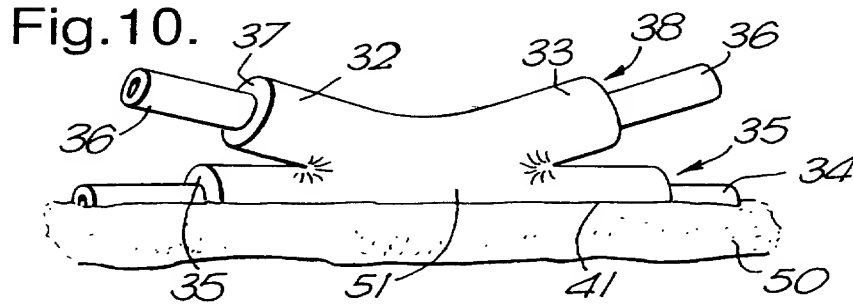
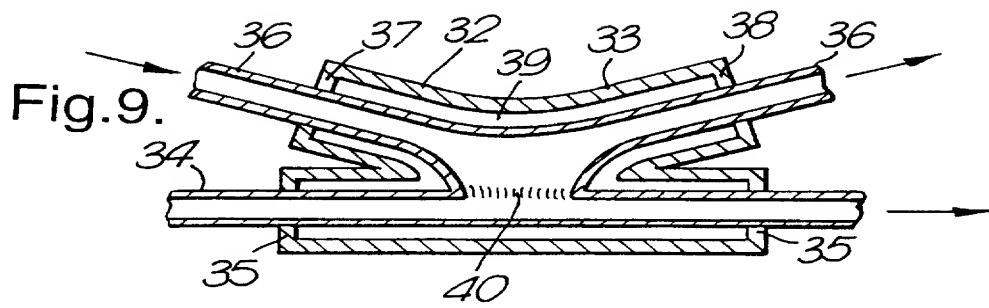
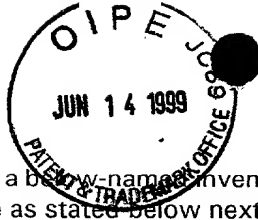


Fig.8.







DECLARATION AND POWER OF ATTORNEY

USA #3
GJE-30

As a below-named inventor, I hereby declare that my residence, post office address and citizenship are as stated below next to my name; I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of subject matter which is claimed and for which a patent is sought on an invention entitled
THERAPEUTIC USE OF AN AGENT THAT STIMULATES NO OR PROSTACYCLIN PRODUCTION AND DELIVERY DEVICE

the specification of which ☐ is attached hereto or

☒ was filed on 03 NOV 1997 as United States Application Number or PCT International Application Number PCT/GB97/03015 and was amended on 16 OCT 1998

(if applicable)
I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above. I acknowledge the duty to disclose information which is material to patentability as defined in 37 CFR 1.56. I hereby claim foreign priority benefits under 35 U.S.C. 119(a)-(d) or 365(b) of any foreign application(s) for patent or inventor's certificate, or 365(a) of any PCT international application which designated at least one country other than the United States of America, listed below and have also identified below, by checking the box, any foreign application for a patent or inventor's certificate, or PCT international application having a filing date before that of the application on which priority is claimed:

Prior Foreign Application Number(s)	Country	Foreign Filing Date	Priority Not Claimed	Certified Copy Attached?	YES	NO
9622852.3	GB	01 NOV 1996	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
9709494.0	GB	09 MAY 1997	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
9717791.9	GB	21 AUG 1997	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

As a named inventor, I hereby appoint the following registered practitioner(s) to prosecute this application and to transact all business in the Patent and Trademark Office connected therewith: David R. Saliwanchik, Reg. 31,794; Jeff Lloyd, Reg. 35,589; Doran R. Pace, Reg. 38,261; Christine O. McLeod, Reg. 36,213; Jay M. Sanders, Reg. 39,355; James S. Parker, Reg. 40,119; Jean E. Kyle, Reg. 36,987 and Timothy H. Van Dyke, Reg. P-43,218.

Direct all correspondence to:
Saliwanchik, Lloyd & Saliwanchik
2421 N.W. 41st Street, Suite A-1
Gainesville, FL 32606-6669
USA

664730" 98446260

8

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under 18 U.S.C 1001 and that such willful false statements may jeopardise the validity of the application or any patent issued thereon.

1-00
Full name of sole or
First Inventor

John Francis MARTIN

Inventor's signature

Residence address

London, United Kingdom. GBX

Post Office address

The Cruciform Project, Saint Martin's House,
Tottenham Court Road, London W1P 9LN, United Kingdom.

Country of Citizenship United Kingdom.

Date of signature 17.5.99

2-00
Full name of
Second Inventor

Seppo YLA-HERTTUALA

Inventor's signature

Residence address

Kuopio, Finland. FIX

Post Office address

A.I. Virtanen Institute, University of Kupio, P.O. Box 1627,
70211 Kuopio, Finland.

Country of Citizenship Finland.

Date of signature 19.5.99

3-00
Full name of
Third Inventor

Stephen George Edward BARKER

Inventor's signature

Residence address

London.

Post Office address

The Vascular Unit, Department of Surgery,
Sir Jules Thorn Building, The Middlesex Hospital,
Mortimer Street, London W1N 8AA.

Country of Citizenship United Kingdom.

Date of signature 17/5/99